

Package ‘GenomicRanges’

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Title Representation and manipulation of genomic intervals

Description The ability to efficiently represent and manipulate genomic annotations and alignments is playing a central role when it comes to analyze high-throughput sequencing data (a.k.a. NGS data). The package defines general purpose containers for storing genomic intervals. Specialized containers for representing and manipulating short alignments against a reference genome are defined in the GenomicAlignments package.

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Collate utils.R phicoef.R transcript-utils.R constraint.R strand-utils.R range-squeezers.R GenomicRanges-class.R GRanges-class.R DelegatingGenomicRanges-class.R GNCList-class.R GIntervalTree-class.R GenomicRanges-comparison.R GenomicRangesList-class.R GRangesList-class.R makeGRangesFromDataFrame.R SummarizedExperiment-class.R

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absoluteRanges

Transform genomic ranges into "absolute" ranges

Description

absoluteRanges transforms the genomic ranges in `x` into *absolute* ranges i.e. into ranges counted from the beginning of the virtual sequence obtained by concatenating all the sequences in the underlying genome (in the order reported by `seqlevels(x)`).

relativeRanges performs the reverse transformation.

NOTE: These functions only work on *small* genomes. See Details section below.

Usage

```
absoluteRanges(x)
relativeRanges(x, seqlengths)
```

```
## Related utility:
isSmallGenome(seqlengths)
```

Arguments

<code>x</code>	For absoluteRanges: a GenomicRanges object with ranges defined on a <i>small</i> genome (see Details section below). For relativeRanges: a Ranges object.
<code>seqlengths</code>	An object holding sequence lengths. This can be a named integer (or numeric) vector with no duplicated names as returned by seqlengths() , or any object from which sequence lengths can be extracted with seqlengths() . For relativeRanges, seqlengths must describe a <i>small</i> genome (see Details section below).

Details

Because absoluteRanges returns the *absolute* ranges in an [IRanges](#) object, and because an [IRanges](#) object cannot hold ranges with an end $> .Machine$integer.max$ (i.e. $\geq 2^{31}$ on most platforms), absoluteRanges cannot be used if the size of the underlying genome (i.e. the total length of the sequences in it) is $> .Machine$integer.max$. Utility function isSmallGenome is provided as a mean for the user to check upfront whether the genome is *small* (i.e. its size is $\leq .Machine$integer.max$) or not, and thus compatible with absoluteRanges or not.

relativeRanges applies the same restriction by looking at the seqlengths argument.

Value

An [IRanges](#) object for absoluteRanges.

A [GRanges](#) object for relativeRanges.

isSmallGenome returns TRUE if the total length of the underlying sequences is $\leq .Machine$integer.max$ (e.g. Fly genome), FALSE if not (e.g. Human genome), or NA if it cannot be computed (because some sequence lengths are NA).

Author(s)

H. Pages

See Also

- [GRanges](#) objects.
- [IRanges](#) objects in the **IRanges** package.
- [Seqinfo](#) objects and the [seqlengths](#) getter in the **GenomeInfoDb** package.
- The [tileGenome](#) function for putting tiles on a genome.

Examples

```
## -----
## TOY EXAMPLE
## -----

gr <- GRanges(Rle(c("chr2", "chr1", "chr3", "chr1"), 4:1),
              IRanges(1:10, width=5),
              seqinfo=Seqinfo(c("chr1", "chr2", "chr3"), c(100, 50, 20)))

ar <- absoluteRanges(gr)
ar

gr2 <- relativeRanges(ar, seqlengths(gr))
gr2

## Sanity check:
stopifnot(all(gr == gr2))

## -----
## ON REAL DATA
## -----

## With a "small" genome

library(TxDb.Dmelanogaster.UCSC.dm3.ensGene)
txdb <- TxDb.Dmelanogaster.UCSC.dm3.ensGene
ex <- exons(txdb)
ex

isSmallGenome(ex)

## Note that because isSmallGenome() can return NA (see Value section
## above), its result should always be wrapped inside isTRUE() when
## used in an if statement:
if (isTRUE(isSmallGenome(ex))) {
  ar <- absoluteRanges(ex)
  ar

  ex2 <- relativeRanges(ar, seqlengths(ex))
  ex2 # original strand is not restored

  ## Sanity check:
  strand(ex2) <- strand(ex) # restore the strand
  stopifnot(all(ex == ex2))
}
```

```

}

## With a "big" genome (but we can reduce it)

library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
ex <- exons(txdb)
isSmallGenome(ex)
## Not run:
  absoluteRanges(ex) # error!

## End(Not run)

## However, if we are only interested in some chromosomes, we might
## still be able to use absoluteRanges():
seqlevels(ex, force=TRUE) <- paste0("chr", 1:10)
isSmallGenome(ex) # TRUE!
ar <- absoluteRanges(ex)
ex2 <- relativeRanges(ar, seqlengths(ex))

## Sanity check:
strand(ex2) <- strand(ex)
stopifnot(all(ex == ex2))

```

Constraints

Enforcing constraints thru Constraint objects

Description

Attaching a Constraint object to an object of class A (the "constrained" object) is meant to be a convenient/reusable/extensible way to enforce a particular set of constraints on particular instances of A.

THIS IS AN EXPERIMENTAL FEATURE AND STILL VERY MUCH A WORK-IN-PROGRESS!

Details

For the developer, using constraints is an alternative to the more traditional approach that consists in creating subclasses of A and implementing specific validity methods for each of them. However, using constraints offers the following advantages over the traditional approach:

- The traditional approach often tends to lead to a proliferation of subclasses of A.
- Constraints can easily be re-used across different classes without the need to create any new class.
- Constraints can easily be combined.

All constraints are implemented as concrete subclasses of the Constraint class, which is a virtual class with no slots. Like the Constraint virtual class itself, concrete Constraint subclasses cannot have slots.

Here are the 7 steps typically involved in the process of putting constraints on objects of class A:

1. Add a slot named `constraint` to the definition of class A. The type of this slot must be `ConstraintORNULL`. Note that any subclass of A will inherit this slot.
2. Implements the `constraint()` accessors (getter and setter) for objects of class A. This is done by implementing the "constraint" method (getter) and replacement method (setter) for objects of class A (see the examples below). As a convenience to the user, the setter should also accept the name of a constraint (i.e. the name of its class) in addition to an instance of that class. Note that those accessors will work on instances of any subclass of A.
3. Modify the validity method for class A so it also returns the result of `checkConstraint(x, constraint(x))` (append this result to the result returned by the validity method).
4. Testing: Create `x`, an instance of class A (or subclass of A). By default there is no constraint on it (`constraint(x)` is `NULL`). `validObject(x)` should return `TRUE`.
5. Create a new constraint (`MyConstraint`) by extending the `Constraint` class, typically with `setClass("MyConstraint", contains="Constraint")`. This constraint is not enforcing anything yet so you could put it on `x` (with `constraint(x) <- "MyConstraint"`), but not much would happen. In order to actually enforce something, a "checkConstraint" method for signature `c(x="A", constraint="MyConstraint")` needs to be implemented.
6. Implement a "checkConstraint" method for signature `c(x="A", constraint="MyConstraint")`. Like validity methods, "checkConstraint" methods must return `NULL` or a character vector describing the problems found. Like validity methods, they should never fail (i.e. they should never raise an error). Note that, alternatively, an existing constraint (e.g. `SomeConstraint`) can be adapted to work on objects of class A by just defining a new "checkConstraint" method for signature `c(x="A", constraint="SomeConstraint")`. Also, stricter constraints can be built on top of existing constraints by extending one or more existing constraints (see the examples below).
7. Testing: Try `constraint(x) <- "MyConstraint"`. It will or will not work depending on whether `x` satisfies the constraint or not. In the former case, trying to modify `x` in a way that breaks the constraint on it will also raise an error.

Note

WARNING: This note is not true anymore as the constraint slot has been temporarily removed from [GenomicRanges](#) objects (starting with package `GenomicRanges` \geq 1.7.9).

Currently, only [GenomicRanges](#) objects can be constrained, that is:

- they have a `constraint` slot;
- they have `constraint()` accessors (getter and setter) for this slot;
- their validity method has been modified so it also returns the result of `checkConstraint(x, constraint(x))`.

More classes in the `GenomicRanges` and `IRanges` packages will support constraints in the near future.

Author(s)

H. Pages

See Also

[setClass](#), [is](#), [setMethod](#), [showMethods](#), [validObject](#), [GenomicRanges-class](#)

Examples

```

## The examples below show how to define and set constraints on
## GenomicRanges objects. Note that this is how the constraint()
## setter is defined for GenomicRanges objects:
#setReplaceMethod("constraint", "GenomicRanges",
#  function(x, value)
#  {
#    if (isSingleString(value))
#      value <- new(value)
#    if (!is(value, "ConstraintORNULL"))
#      stop("the supplied 'constraint' must be a ",
#           "Constraint object, a single string, or NULL")
#    x@constraint <- value
#    validObject(x)
#    x
#  }
#)

#selectMethod("constraint", "GenomicRanges") # the getter
#selectMethod("constraint<-", "GenomicRanges") # the setter

## We'll use the GRanges instance 'gr' created in the GRanges examples
## to test our constraints:
example(GRanges, echo=FALSE)
gr
#constraint(gr)

## -----
## EXAMPLE 1: The HasRangeTypeCol constraint.
## -----
## The HasRangeTypeCol constraint checks that the constrained object
## has a unique "rangeType" metadata column and that this column
## is a 'factor' Rle with no NAs and with the following levels
## (in this order): gene, transcript, exon, cds, 5utr, 3utr.

setClass("HasRangeTypeCol", contains="Constraint")

## Like validity methods, "checkConstraint" methods must return NULL or
## a character vector describing the problems found. They should never
## fail i.e. they should never raise an error.
setMethod("checkConstraint", c("GenomicRanges", "HasRangeTypeCol"),
  function(x, constraint, verbose=FALSE)
  {
    x_mcols <- mcols(x)
    idx <- match("rangeType", colnames(x_mcols))
    if (length(idx) != 1L || is.na(idx)) {
      msg <- c("'mcols(x)' must have exactly 1 column ",
               "named \"rangeType\"")
      return(paste(msg, collapse=""))
    }
    rangeType <- x_mcols[[idx]]
    .LEVELS <- c("gene", "transcript", "exon", "cds", "5utr", "3utr")
  }

```

```

    if (!is(rangeType, "Rle") ||
        S4Vectors:::anyMissing(runValue(rangeType)) ||
        !identical(levels(rangeType), .LEVELS))
    {
      msg <- c("'mcols(x)$rangeType' must be a ",
              "'factor' Rle with no NAs and with levels: ",
              paste(.LEVELS, collapse=", "))
      return(paste(msg, collapse=""))
    }
    NULL
  }
)

#\dontrun{
#constraint(gr) <- "HasRangeTypeCol" # will fail
#}
checkConstraint(gr, new("HasRangeTypeCol")) # with GenomicRanges >= 1.7.9

levels <- c("gene", "transcript", "exon", "cds", "5utr", "3utr")
rangeType <- Rle(factor(c("cds", "gene"), levels=levels), c(8, 2))
mcols(gr)$rangeType <- rangeType
#constraint(gr) <- "HasRangeTypeCol" # OK
checkConstraint(gr, new("HasRangeTypeCol")) # with GenomicRanges >= 1.7.9

## Use is() to check whether the object has a given constraint or not:
#is(constraint(gr), "HasRangeTypeCol") # TRUE
#\dontrun{
#mcols(gr)$rangeType[3] <- NA # will fail
#}
mcols(gr)$rangeType[3] <- NA
checkConstraint(gr, new("HasRangeTypeCol")) # with GenomicRanges >= 1.7.9

## -----
## EXAMPLE 2: The GeneRanges constraint.
## -----
## The GeneRanges constraint is defined on top of the HasRangeTypeCol
## constraint. It checks that all the ranges in the object are of type
## "gene".

setClass("GeneRanges", contains="HasRangeTypeCol")

## The checkConstraint() generic will check the HasRangeTypeCol constraint
## first, and, only if it's satisfied, it will then check the GeneRanges
## constraint.
setMethod("checkConstraint", c("GenomicRanges", "GeneRanges"),
  function(x, constraint, verbose=FALSE)
  {
    rangeType <- mcols(x)$rangeType
    if (!all(rangeType == "gene")) {
      msg <- c("all elements in 'mcols(x)$rangeType' ",
              "must be equal to \"gene\"")
      return(paste(msg, collapse=""))
    }
  }
)

```



```

        NULL
    }
)

#\dontrun{
#constraint(gr) <- "GeneRanges" # will fail
#}
checkConstraint(gr, new("GeneRanges")) # with GenomicRanges >= 1.7.9

mcols(gr)$rangeType[] <- "gene"
## This replace the previous constraint (HasRangeTypeCol):
#constraint(gr) <- "GeneRanges" # OK
checkConstraint(gr, new("GeneRanges")) # with GenomicRanges >= 1.7.9

#is(constraint(gr), "GeneRanges") # TRUE
## However, 'gr' still indirectly has the HasRangeTypeCol constraint
## (because the GeneRanges constraint extends the HasRangeTypeCol
## constraint):
#is(constraint(gr), "HasRangeTypeCol") # TRUE
#\dontrun{
#mcols(gr)$rangeType[] <- "exon" # will fail
#}
mcols(gr)$rangeType[] <- "exon"
checkConstraint(gr, new("GeneRanges")) # with GenomicRanges >= 1.7.9

## -----
## EXAMPLE 3: The HasGCCol constraint.
## -----
## The HasGCCol constraint checks that the constrained object has a
## unique "GC" metadata column, that this column is of type numeric,
## with no NAs, and that all the values in that column are >= 0 and <= 1.

setClass("HasGCCol", contains="Constraint")

setMethod("checkConstraint", c("GenomicRanges", "HasGCCol"),
  function(x, constraint, verbose=FALSE)
  {
    x_mcols <- mcols(x)
    idx <- match("GC", colnames(x_mcols))
    if (length(idx) != 1L || is.na(idx)) {
      msg <- c("'mcols(x)' must have exactly ",
              "one column named \"GC\"")
      return(paste(msg, collapse=""))
    }
    GC <- x_mcols[[idx]]
    if (!is.numeric(GC) ||
        S4Vectors:::anyMissing(GC) ||
        any(GC < 0) || any(GC > 1))
    {
      msg <- c("'mcols(x)$GC' must be a numeric vector ",
              "with no NAs and with values between 0 and 1")
      return(paste(msg, collapse=""))
    }
  }
}

```

```

    NULL
  }
)

## This replace the previous constraint (GeneRanges):
#constraint(gr) <- "HasGCCol" # OK
checkConstraint(gr, new("HasGCCol")) # with GenomicRanges >= 1.7.9

#is(constraint(gr), "HasGCCol") # TRUE
#is(constraint(gr), "GeneRanges") # FALSE
#is(constraint(gr), "HasRangeTypeCol") # FALSE

## -----
## EXAMPLE 4: The HighGCRanges constraint.
## -----
## The HighGCRanges constraint is defined on top of the HasGCCol
## constraint. It checks that all the ranges in the object have a GC
## content >= 0.5.

setClass("HighGCRanges", contains="HasGCCol")

## The checkConstraint() generic will check the HasGCCol constraint
## first, and, if it's statisfied, it will then check the HighGCRanges
## constraint.
setMethod("checkConstraint", c("GenomicRanges", "HighGCRanges"),
  function(x, constraint, verbose=FALSE)
  {
    GC <- mcols(x)$GC
    if (!all(GC >= 0.5)) {
      msg <- c("all elements in 'mcols(x)$GC' ",
              "must be >= 0.5")
      return(paste(msg, collapse=""))
    }
    NULL
  }
)

#\dontrun{
#constraint(gr) <- "HighGCRanges" # will fail
#}
checkConstraint(gr, new("HighGCRanges")) # with GenomicRanges >= 1.7.9
mcols(gr)$GC[6:10] <- 0.5
#constraint(gr) <- "HighGCRanges" # OK
checkConstraint(gr, new("HighGCRanges")) # with GenomicRanges >= 1.7.9

#is(constraint(gr), "HighGCRanges") # TRUE
#is(constraint(gr), "HasGCCol") # TRUE

## -----
## EXAMPLE 5: The HighGCCGeneRanges constraint.
## -----
## The HighGCCGeneRanges constraint is the combination (AND) of the
## GeneRanges and HighGCRanges constraints.

```

```

setClass("HighGCCGeneRanges", contains=c("GeneRanges", "HighGCRanges"))

## No need to define a method for this constraint: the checkConstraint()
## generic will automatically check the GeneRanges and HighGCRanges
## constraints.

#constraint(gr) <- "HighGCCGeneRanges" # OK
checkConstraint(gr, new("HighGCCGeneRanges")) # with GenomicRanges >= 1.7.9

#is(constraint(gr), "HighGCCGeneRanges") # TRUE
#is(constraint(gr), "HighGCRanges") # TRUE
#is(constraint(gr), "HasGCCol") # TRUE
#is(constraint(gr), "GeneRanges") # TRUE
#is(constraint(gr), "HasRangeTypeCol") # TRUE

## See how all the individual constraints are checked (from less
## specific to more specific constraints):
#checkConstraint(gr, constraint(gr), verbose=TRUE)
checkConstraint(gr, new("HighGCCGeneRanges"), verbose=TRUE) # with
# GenomicRanges
# >= 1.7.9

## See all the "checkConstraint" methods:
showMethods("checkConstraint")

```

coverage-methods

Coverage of a GRanges or GRangesList object

Description

[coverage](#) methods for [GRanges](#) and [GRangesList](#) objects.

NOTE: The [coverage](#) generic function and methods for [Ranges](#) and [RangesList](#) objects are defined and documented in the **IRanges** package. Methods for [GAlignments](#) and [GAlignmentPairs](#) objects are defined and documented in the **GenomicAlignments** package.

Usage

```

## S4 method for signature 'GenomicRanges'
coverage(x, shift=0L, width=NULL, weight=1L,
         method=c("auto", "sort", "hash"))

## S4 method for signature 'GRangesList'
coverage(x, shift=0L, width=NULL, weight=1L,
         method=c("auto", "sort", "hash"))

```

Arguments

x	A GRanges or GRangesList object.
shift	<p>A numeric vector or a list-like object. If numeric, it must be parallel to x (recycled if necessary). If a list-like object, it must have 1 list element per seqlevel in x, and its names must be exactly <code>seqlevels(x)</code>.</p> <p>Alternatively, shift can also be specified as a single string naming a metadata column in x (i.e. a column in <code>mcols(x)</code>) to be used as the shift vector.</p> <p>See ?coverage in the IRanges package for more information about this argument.</p>
width	<p>Either NULL (the default), or an integer vector. If NULL, it is replaced with <code>seqlengths(x)</code>. Otherwise, the vector must have the length and names of <code>seqlengths(x)</code> and contain NAs or non-negative integers.</p> <p>See ?coverage in the IRanges package for more information about this argument.</p>
weight	<p>A numeric vector or a list-like object. If numeric, it must be parallel to x (recycled if necessary). If a list-like object, it must have 1 list element per seqlevel in x, and its names must be exactly <code>seqlevels(x)</code>.</p> <p>Alternatively, weight can also be specified as a single string naming a metadata column in x (i.e. a column in <code>mcols(x)</code>) to be used as the weight vector.</p> <p>See ?coverage in the IRanges package for more information about this argument.</p>
method	See ?coverage in the IRanges package for a description of this argument.

Details

When x is a [GRangesList](#) object, `coverage(x, ...)` is equivalent to `coverage(unlist(x), ...)`.

Value

A named [RleList](#) object with one coverage vector per seqlevel in x.

Author(s)

H. Pages and P. Aboyoun

See Also

- [coverage](#) in the **IRanges** package.
- [coverage-methods](#) in the **GenomicAlignments** package.
- [RleList](#) objects in the **IRanges** package.
- [GRanges](#) and [GRangesList](#) objects.

Examples

```
## Coverage of a GRanges object:
gr <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "* ", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
cvg <- coverage(gr)
pcvg <- coverage(gr[strand(gr) == "+"])
mcvg <- coverage(gr[strand(gr) == "-"])
scvg <- coverage(gr[strand(gr) == "*"])
stopifnot(identical(pcvg + mcvg + scvg, cvg))

## Coverage of a GRangesList object:
gr1 <- GRanges(seqnames="chr2",
  ranges=IRanges(3, 6),
  strand = "+")
gr2 <- GRanges(seqnames=c("chr1", "chr1"),
  ranges=IRanges(c(7,13), width=3),
  strand=c("+", "-"))
gr3 <- GRanges(seqnames=c("chr1", "chr2"),
  ranges=IRanges(c(1, 4), c(3, 9)),
  strand=c("-", "-"))
grl <- GRangesList(gr1=gr1, gr2=gr2, gr3=gr3)
stopifnot(identical(coverage(grl), coverage(unlist(grl))))
```

DelegatingGenomicRanges-class

DelegatingGenomicRanges objects

Description

The DelegatingGenomicRanges class implements the virtual GenomicRanges class using a delegate GenomicRanges. This enables developers to create GenomicRanges subclasses that add specialized columns or other structure, while remaining agnostic to how the data are actually stored. See the Extending GenomicRanges vignette.

Author(s)

M. Lawrence

 findOverlaps-methods *Finding overlapping genomic ranges*

Description

Various methods for finding/counting overlaps between objects containing genomic ranges. This man page describes the methods that operate on [GenomicRanges](#) or [GRangesList](#) objects.

NOTE: The [findOverlaps](#) generic function and methods for [Ranges](#) and [RangesList](#) objects are defined and documented in the **IRanges** package. The methods for [GAlignments](#), [GAlignmentPairs](#), and [GAlignmentsList](#) objects are defined and documented in the **GenomicAlignments** package.

Usage

```
## S4 method for signature 'GenomicRanges,GenomicRanges'
findOverlaps(query, subject,
             maxgap=0L, minoverlap=1L,
             type=c("any", "start", "end", "within", "equal"),
             select=c("all", "first", "last", "arbitrary"),
             algorithm=c("nclist", "intervaltree"),
             ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges,GenomicRanges'
countOverlaps(query, subject,
             maxgap=0L, minoverlap=1L,
             type=c("any", "start", "end", "within", "equal"),
             algorithm=c("nclist", "intervaltree"),
             ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges,GenomicRanges'
overlapsAny(query, subject,
            maxgap=0L, minoverlap=1L,
            type=c("any", "start", "end", "within", "equal"),
            algorithm=c("nclist", "intervaltree"),
            ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges,GenomicRanges'
subsetByOverlaps(query, subject,
                 maxgap=0L, minoverlap=1L,
                 type=c("any", "start", "end", "within", "equal"),
                 algorithm=c("nclist", "intervaltree"),
                 ignore.strand=FALSE)
```

Arguments

query, subject A [GRanges](#) or [GRangesList](#) object. [RangesList](#) and [RangedData](#) are also accepted for one of query or subject.

maxgap, minoverlap, type, algorithm	See findOverlaps in the IRanges package for a description of these arguments.
select	When select is "all" (the default), the results are returned as a Hits object. Otherwise the returned value is an integer vector parallel to query (i.e. same length) containing the first, last, or arbitrary overlapping interval in subject, with NA indicating intervals that did not overlap any intervals in subject.
ignore.strand	When set to TRUE, the strand information is ignored in the overlap calculations.

Details

When the query and the subject are [GRanges](#) or [GRangesList](#) objects, `findOverlaps` uses the triplet (sequence name, range, strand) to determine which features (see paragraph below for the definition of feature) from the query overlap which features in the subject, where a strand value of "*" is treated as occurring on both the "+" and "-" strand. An overlap is recorded when a feature in the query and a feature in the subject have the same sequence name, have a compatible pairing of strands (e.g. "+"/"+", "-"/"-", "*"/"+", "*"/"-", etc.), and satisfy the interval overlap requirements. Strand is taken as "*" for [RangedData](#) and [RangesList](#).

In the context of `findOverlaps`, a feature is a collection of ranges that are treated as a single entity. For [GRanges](#) objects, a feature is a single range; while for [GRangesList](#) objects, a feature is a list element containing a set of ranges. In the results, the features are referred to by number, which run from 1 to `length(query)/length(subject)`.

Value

For `findOverlaps` either a [Hits](#) object when `select="all"` or an integer vector otherwise.

For `countOverlaps` an integer vector containing the tabulated query overlap hits.

For `overlapsAny` a logical vector of length equal to the number of ranges in query indicating those that overlap any of the ranges in subject.

For `subsetByOverlaps` an object of the same class as query containing the subset that overlapped at least one entity in subject.

For [RangedData](#) and [RangesList](#), with the exception of `subsetByOverlaps`, the results align to the unlisted form of the object. This turns out to be fairly convenient for [RangedData](#) (not so much for [RangesList](#), but something has to give).

Author(s)

P. Aboyoun, S. Falcon, M. Lawrence, and H. Pages

See Also

- The [Hits](#) class for representing a set of hits between 2 vector-like objects.
- The [findOverlaps](#) generic function defined in the **IRanges** package.
- The [GNCList](#) constructor and class for preprocessing and representing a [GenomicRanges](#) or object as a data structure based on Nested Containment Lists.
- The [GRanges](#) and [GRangesList](#) classes.

Examples

```

## -----
## BASIC EXAMPLES
## -----

## GRanges object:
gr <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, width=10:1, names=head(letters,10)),
  strand=Rle(strand(c("-", "+", "*+", "+", "-")), c(1, 2, 2, 3, 2)),
  score=1:10,
  GC=seq(1, 0, length=10)
)

gr

## GRangesList object:
gr1 <- GRanges(seqnames="chr2", ranges=IRanges(4:3, 6),
  strand="+", score=5:4, GC=0.45)
gr2 <- GRanges(seqnames=c("chr1", "chr1"),
  ranges=IRanges(c(7,13), width=3),
  strand=c("+", "-"), score=3:4, GC=c(0.3, 0.5))
gr3 <- GRanges(seqnames=c("chr1", "chr2"),
  ranges=IRanges(c(1, 4), c(3, 9)),
  strand=c("-", "-"), score=c(6L, 2L), GC=c(0.4, 0.1))
grl <- GRangesList("gr1"=gr1, "gr2"=gr2, "gr3"=gr3)

## Overlapping two GRanges objects:
table(!is.na(findOverlaps(gr, gr1, select="arbitrary")))
countOverlaps(gr, gr1)
findOverlaps(gr, gr1)
subsetByOverlaps(gr, gr1)

countOverlaps(gr, gr1, type="start")
findOverlaps(gr, gr1, type="start")
subsetByOverlaps(gr, gr1, type="start")

findOverlaps(gr, gr1, select="first")
findOverlaps(gr, gr1, select="last")

findOverlaps(gr1, gr)
findOverlaps(gr1, gr, type="start")
findOverlaps(gr1, gr, type="within")
findOverlaps(gr1, gr, type="equal")

## -----
## MORE EXAMPLES
## -----

table(!is.na(findOverlaps(gr, gr1, select="arbitrary")))
countOverlaps(gr, gr1)
findOverlaps(gr, gr1)
subsetByOverlaps(gr, gr1)

```



```
## Overlaps between a GRanges and a GRangesList object:

table(!is.na(findOverlaps(grl, gr, select="first")))
countOverlaps(grl, gr)
findOverlaps(grl, gr)
subsetByOverlaps(grl, gr)
countOverlaps(grl, gr, type="start")
findOverlaps(grl, gr, type="start")
subsetByOverlaps(grl, gr, type="start")
findOverlaps(grl, gr, select="first")

table(!is.na(findOverlaps(grl, gr1, select="first")))
countOverlaps(grl, gr1)
findOverlaps(grl, gr1)
subsetByOverlaps(grl, gr1)
countOverlaps(grl, gr1, type="start")
findOverlaps(grl, gr1, type="start")
subsetByOverlaps(grl, gr1, type="start")
findOverlaps(grl, gr1, select="first")

## Overlaps between two GRangesList objects:
countOverlaps(grl, rev(gr1))
findOverlaps(grl, rev(gr1))
subsetByOverlaps(grl, rev(gr1))
```

GenomicRanges-comparison

Comparing and ordering genomic ranges

Description

Methods for comparing and ordering the elements in one or more [GenomicRanges](#) objects.

Usage

```
## duplicated()
## -----

## S4 method for signature 'GenomicRanges'
duplicated(x, incomparables=FALSE, fromLast=FALSE,
           method=c("auto", "quick", "hash"))

## match()
## -----

## S4 method for signature 'GenomicRanges,GenomicRanges'
match(x, table, nomatch=NA_integer_, incomparables=NULL,
      method=c("auto", "quick", "hash"), ignore.strand=FALSE)
```

```

## order() and related methods
## -----

## S4 method for signature 'GenomicRanges'
order(..., na.last=TRUE, decreasing=FALSE)

## S4 method for signature 'GenomicRanges'
sort(x, decreasing=FALSE, ignore.strand=FALSE, by)

## S4 method for signature 'GenomicRanges'
rank(x, na.last=TRUE,
      ties.method=c("average", "first", "random", "max", "min"))

## Generalized element-wise (aka "parallel") comparison of 2 GenomicRanges
## objects
## -----

## S4 method for signature 'GenomicRanges,GenomicRanges'
compare(x, y)

```

Arguments

<code>x</code> , <code>table</code> , <code>y</code>	GenomicRanges objects.
<code>incomparables</code>	Not supported.
<code>fromLast</code> , <code>method</code> , <code>nomatch</code>	See <code>?`Ranges-comparison`</code> in the <code>IRanges</code> package for a description of these arguments.
<code>ignore.strand</code>	Whether or not the strand should be ignored when comparing 2 genomic ranges.
<code>...</code>	Additional GenomicRanges objects used for breaking ties.
<code>na.last</code>	Ignored.
<code>decreasing</code>	TRUE or FALSE.
<code>ties.method</code>	A character string specifying how ties are treated. Only "first" is supported for now.
<code>by</code>	An optional formula that is resolved against <code>as.env(x)</code> ; the resulting variables are passed to <code>order</code> to generate the ordering permutation.

Details

Two elements of a [GenomicRanges](#) object (i.e. two genomic ranges) are considered equal iff they are on the same underlying sequence and strand, and have the same start and width. `duplicated()` and `unique()` on a [GenomicRanges](#) object are conforming to this.

The "natural order" for the elements of a [GenomicRanges](#) object is to order them (a) first by sequence level, (b) then by strand, (c) then by start, (d) and finally by width. This way, the space of genomic ranges is totally ordered. Note that the `reduce` method for [GenomicRanges](#) uses this "natural order" implicitly. Also, note that, because we already do (c) and (d) for regular ranges (see

?`Ranges-comparison`), genomic ranges that belong to the same underlying sequence and strand are ordered like regular ranges. `order()`, `sort()`, and `rank()` on a `GenomicRanges` object are using this "natural order".

Also `==`, `!=`, `<=`, `>=`, `<` and `>` on `GenomicRanges` objects are using this "natural order".

Author(s)

H. Pages

See Also

- The `GenomicRanges` class.
- `Ranges-comparison` in the `IRanges` package for comparing and ordering genomic ranges.
- `intra-range-methods` and `inter-range-methods` for intra and inter range transformations.
- `setops-methods` for set operations on `GenomicRanges` objects.
- `findOverlaps-methods` for finding overlapping genomic ranges.

Examples

```
gr0 <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(c(1:9,7L), end=10),
  strand=Rle(strand(c("-", "+", "*"), "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
gr <- c(gr0, gr0[7:3])
names(gr) <- LETTERS[seq_along(gr)]

## -----
## A. ELEMENT-WISE (AKA "PARALLEL") COMPARISON OF 2 GenomicRanges OBJECTS
## -----
gr[2] == gr[2] # TRUE
gr[2] == gr[5] # FALSE
gr == gr[4]
gr >= gr[3]

## -----
## B. duplicated(), unique()
## -----
duplicated(gr)
unique(gr)

## -----
## C. match(), %in%
## -----
table <- gr[1:7]
match(gr, table)
match(gr, table, ignore.strand=TRUE)

gr %in% table
```

```

## -----
## D. findMatches(), countMatches()
## -----
findMatches(gr, table)
countMatches(gr, table)

findMatches(gr, table, ignore.strand=TRUE)
countMatches(gr, table, ignore.strand=TRUE)

gr_levels <- unique(gr)
countMatches(gr_levels, gr)

## -----
## E. order() AND RELATED METHODS
## -----
order(gr)
sort(gr)
sort(gr, ignore.strand=TRUE)

## TODO: Broken. Please fix!
#sort(gr, by = ~ seqnames + start + end) # equivalent to (but slower than) above

score(gr) <- rev(seq_len(length(gr)))

## TODO: Broken. Please fix!
#sort(gr, by = ~ score)

rank(gr)

## -----
## F. GENERALIZED ELEMENT-WISE COMPARISON OF 2 GenomicRanges OBJECTS
## -----
gr2 <- GRanges(c(rep("chr1", 12), "chr2"), IRanges(c(1:11, 6:7), width=3))
strand(gr2)[12] <- "+"
gr3 <- GRanges("chr1", IRanges(5, 9))

compare(gr2, gr3)
rangeComparisonCodeToLetter(compare(gr2, gr3))

```

GenomicRangesList-class

GenomicRangesList objects

Description

A `GenomicRangesList` is a [List](#) of `GenomicRanges`. It is a virtual class; `SimpleGenomicRangesList` is the basic implementation. The subclass `GRangesList` provides special behavior and is particularly efficient for storing a large number of elements.

Constructor

`GenomicRangesList(...)`: Constructs a `SimpleGenomicRangesList` with elements taken from the arguments in `...`. If the only argument is a list, the elements are taken from that list.

Coercion

`as(from, "GenomicRangesList")`: Supported from types include:

RangedDataList Each element of `from` is coerced to a `GenomicRanges`.

`as(from, "RangedDataList")`: Supported from types include:

GenomicRangesList Each element of `from` is coerced to a `RangedData`.

Author(s)

Michael Lawrence

See Also

[GRangesList](#), which differs from `SimpleGenomicRangesList` in that the `GRangesList` treats its elements as single, compound ranges, particularly in overlap operations. `SimpleGenomicRangesList` is just a barebones list for now, without that compound semantic.

GIntervalTree-class *GIntervalTree* objects

Description

The `GIntervalTree` class is a container for the genomic locations and their associated annotations.

WARNING: `GIntervalTree` objects are deprecated. Please use [GNCList](#) objects instead. See [?GNCList](#) for more information.

Details

A common type of query that arises when working with intervals is finding which intervals in one set overlap those in another. An efficient family of algorithms for answering such queries is known as the Interval Tree. The `GIntervalTree` class implements persistent Interval Trees for efficient querying of genomic intervals. It uses the [IntervalForest](#) class to store a set of trees, one for each `seqlevel` in a [GRanges](#) object.

The simplest approach for finding overlaps is to call the [findOverlaps](#) function on a [Ranges](#) or other object with range information. See the man page of [findOverlaps](#) for how to use this and other related functions. A `GIntervalTree` object is a derivative of [GenomicRanges](#) and stores its genomic ranges as a set of trees (a forest, with one tree per `seqlevel`) that is optimized for overlap queries. Thus, for repeated queries against the same subject, it is more efficient to create a `GIntervalTree` once for the subject using the constructor described below and then perform the queries against the `GIntervalTree` instance.

Like its [GenomicRanges](#) parent class, the `GIntervalTree` class stores the sequences of genomic locations and associated annotations. Each element in the sequence is comprised of a sequence name, an interval, a [strand](#), and optional metadata columns (e.g. score, GC content, etc.). This information is stored in four components as in [GenomicRanges](#), but two of these components are treated in a specific way:

`ranges` an [IntervalForest](#) object containing the ranges stored as a set of interval trees.

`seqnames` these are not stored directly in this class, but are obtained from the `partitioning` component of the [IntervalForest](#) object stored in `ranges`.

Note that `GIntervalTree` objects are not supported for [GRanges](#) objects with circular genomes.

Constructor

`GIntervalTree(x)`: Creates a `GIntervalTree` object from a [GRanges](#) object.

`x` a [GRanges](#) object containing the genomic ranges.

Coercion

`as(from, "GIntervalTree")`: Creates a `GIntervalTree` object from a [GRanges](#) object. `as(from, "GRanges")`: Creates a [GRanges](#) object from an `GIntervalTree` object

Accessors

In the following code snippets, `x` is a `GIntervalTree` object.

`length(x)`: Get the number of elements.

`seqnames(x)`: Get the sequence names.

`ranges(x)`: Get the ranges as an [IRanges](#) object. This is for consistency with the `ranges` accessor for [GRanges](#) objects. To access the underlying [IntervalForest](#) object use the `obj@ranges` form.

`strand(x)`: Get the strand.

`mcols(x, use.names=FALSE)`, `mcols(x) <- value`: Get or set the metadata columns. If `use.names=TRUE` and the metadata columns are not `NULL`, then the names of `x` are propagated as the row names of the returned [DataFrame](#) object. When setting the metadata columns, the supplied value must be `NULL` or a `data.frame`-like object (i.e. [DataTable](#) or `data.frame`) object holding element-wise metadata.

`elementMetadata(x)`, `elementMetadata(x) <- value`, `values(x)`, `values(x) <- value`: Alternatives to `mcols` functions. Their use is discouraged.

`seqinfo(x)`: Get or set the information about the underlying sequences. `value` must be a [Seqinfo](#) object.

`seqlevels(x)`: Get the sequence levels. These are stored in the `partition` slot of the underlying [IntervalForest](#) object.

`seqlengths(x)`: Get the sequence lengths.

`isCircular(x)`: Get the circularity flags. Note that `GIntervalTree` objects are not supported for circular genomes.

`genome(x)`: Get or the genome identifier or assembly name for each sequence.

seqlevelsStyle(x): Get the seqname style for x. See the [seqlevelsStyle](#) generic getter in the **GenomeInfoDb** package for more information.

score(x): Get the “score” column from the element metadata, if any.

Ranges methods

In the following code snippets, x is a GIntervalTree object.

start(x): Get start(ranges(x)).

end(x): Get end(ranges(x)).

width(x): Get width(ranges(x)).

Subsetting

In the code snippets below, x is a GIntervalTree object.

x[i, j]: Get elements i with optional metadata columns mcols(x)[, j], where i can be missing; an NA-free logical, numeric, or character vector; or a 'logical' Rle object.

Other methods

show(x): By default the show method displays 5 head and 5 tail elements. This can be changed by setting the global options showHeadLines and showTailLines. If the object length is less than (or equal to) the sum of these 2 options plus 1, then the full object is displayed.

Author(s)

Hector Corrada Bravo, P. Aboyoun

See Also

[seqinfo](#), [IntervalForest](#), [IntervalTree](#), [findOverlaps-methods](#),

Examples

```
if (interactive()) {
  ## GIntervalTree objects are deprecated. Please use GNCList objects
  ## instead. See ?GNCList for more information.
  seqinfo <- Seqinfo(paste0("chr", 1:3), c(1000, 2000, 1500), NA, "mock1")
  gr <-
    GRanges(seqnames =
      Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
      ranges = IRanges(
        1:10, width = 10:1, names = head(letters,10)),
      strand = Rle(
        strand(c("-", "+", "*+", "+", "-")),
        c(1, 2, 2, 3, 2)),
      score = 1:10,
      GC = seq(1, 0, length=10),
      seqinfo=seqinfo)
  tree <- GIntervalTree(gr)
```

```

tree

## Summarizing elements
table(seqnames(tree))
sum(width(tree))
summary(mcols(tree)[,"score"])

## find Overlaps
subject <-
  GRanges(seqnames =
    Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
    ranges =
    IRanges(1:10, width = 10:1, names = head(letters,10)),
    strand =
    Rle(strand(c("-", "+", "*", "+", "-")),
      c(1, 2, 2, 3, 2)),
    score = 1:10,
    GC = seq(1, 0, length=10))
query <-
  GRanges(seqnames = "chr2", ranges = IRanges(4:3, 6),
    strand = "+", score = 5:4, GC = 0.45)

stree <- GIntervalTree(subject)
findOverlaps(query, stree)
countOverlaps(query, stree)
}

```

GNCList-class

GNCList objects

Description

The GNCList class is a container for storing the Nested Containment List representation of a vector of genomic ranges (typically represented as a [GRanges](#) object). To preprocess a [GRanges](#) object, simply call the `GNCList` constructor function on it. The resulting GNCList object can then be used for efficient overlap-based operations on the genomic ranges.

Usage

```
GNCList(x)
```

Arguments

`x` The [GRanges](#) (or more generally [GenomicRanges](#)) object to preprocess.

Details

The [IRanges](#) package also defines the [NCList](#) and [NCLists](#) constructors and classes for preprocessing and representing a [Ranges](#) or [RangesList](#) object as a data structure based on Nested Containment Lists.

Note that GNCList objects are replacements for [GIntervalTree](#) objects. The latter are deprecated starting with BioC 3.1.

See [?NCList](#) in the **IRanges** package for some important differences between the new algorithm based on Nested Containment Lists and the old algorithm based on interval trees. In particular, the new algorithm supports preprocessing of a [GenomicRanges](#) object with ranges defined on circular sequences (e.g. on the mitochondrial chromosome). See below for some examples.

Value

A GNCList object.

Author(s)

H. Pages

References

Alexander V. Alekseyenko and Christopher J. Lee – Nested Containment List (NCList): a new algorithm for accelerating interval query of genome alignment and interval databases. *Bioinformatics* (2007) 23 (11): 1386-1393. doi: 10.1093/bioinformatics/btl647

See Also

- The [NCList](#) and [NCLists](#) constructors and classes defined in the **IRanges** package.
- [findOverlaps](#) for finding/counting interval overlaps between two *range-based* objects.
- [GRanges](#) objects.

Examples

```
## The examples below are for illustration purpose only and do NOT
## reflect typical usage. This is because, for a one time use, it is
## NOT advised to explicitly preprocess the input for findOverlaps()
## or countOverlaps(). These functions will take care of it and do a
## better job at it (by preprocessing only what's needed when it's
## needed, and release memory as they go).

## -----
## PREPROCESS QUERY OR SUBJECT
## -----

query <- GRanges(Rle(c("chrM", "chr1", "chrM", "chr1"), 4:1),
                 IRanges(1:10, width=5), strand=rep(c("+", "-"), 5))
subject <- GRanges(Rle(c("chr1", "chr2", "chrM"), 3:1),
                  IRanges(6:1, width=5), strand="+")

## Either the query or the subject of findOverlaps() can be preprocessed:

ppsubject <- GNCList(subject)
hits1a <- findOverlaps(query, ppsubject)
hits1a
```

```

hits1b <- findOverlaps(query, ppsubject, ignore.strand=TRUE)
hits1b

ppquery <- GNCList(query)
hits2a <- findOverlaps(ppquery, subject)
hits2a
hits2b <- findOverlaps(ppquery, subject, ignore.strand=TRUE)
hits2b

## Note that 'hits1a' and 'hits2a' contain the same hits but not
## necessarily in the same order.
stopifnot(identical(sort(hits1a), sort(hits2a)))
## Same for 'hits1b' and 'hits2b'.
stopifnot(identical(sort(hits1b), sort(hits2b)))

## -----
## WITH CIRCULAR SEQUENCES
## -----

seqinfo <- Seqinfo(c("chr1", "chr2", "chrM"),
                  seqlengths=c(100, 50, 10),
                  isCircular=c(FALSE, FALSE, TRUE))
seqinfo(query) <- seqinfo[seqlevels(query)]
seqinfo(subject) <- seqinfo[seqlevels(subject)]

ppsubject <- GNCList(subject)
hits3 <- findOverlaps(query, ppsubject)
hits3

## Circularity introduces more hits:

stopifnot(all(hits1a %in% hits3))
new_hits <- setdiff(hits3, hits1a)
new_hits # 1 new hit
query[queryHits(new_hits)]
subject[subjectHits(new_hits)] # positions 11:13 on chrM are the same
                               # as positions 1:3

## Sanity checks:
stopifnot(identical(new_hits, Hits(9, 6, 10, 6)))
ppquery <- GNCList(query)
hits4 <- findOverlaps(ppquery, subject)
stopifnot(identical(sort(hits3), sort(hits4)))

```

GRanges-class

GRanges objects

Description

The GRanges class is a container for the genomic locations and their associated annotations.

Details

GRanges is a vector of genomic locations and associated annotations. Each element in the vector is comprised of a sequence name, an interval, a [strand](#), and optional metadata columns (e.g. score, GC content, etc.). This information is stored in four components:

seqnames a 'factor' [Rle](#) object containing the sequence names.

ranges an [IRanges](#) object containing the ranges.

strand a 'factor' [Rle](#) object containing the [strand](#) information.

mcols a [DataFrame](#) object containing the metadata columns. Columns cannot be named "seqnames", "ranges", "strand", "seqlevels", "seqlengths", "isCircular", "start", "end", "width", or "element".

seqinfo a [Seqinfo](#) object containing information about the set of genomic sequences present in the GRanges object.

Constructor

GRanges(seqnames=Rle(), ranges=NULL, strand=NULL, ..., seqlengths=NULL)
Creates a GRanges object.

seqnames An [Rle](#) object, character vector, or factor containing the sequence names.

ranges An [IRanges](#) object containing the ranges.

strand [Rle](#) object, character vector, or factor containing the strand information.

... Optional metadata columns. These columns cannot be named "start", "end", "width", or "element".

seqlengths An integer vector named with levels(seqnames) and containing the lengths (or NA) for each level in levels(seqnames).

seqinfo A [Seqinfo](#) object containing allowed sequence names, lengths (or NA), and circularity flag, for each level in levels(seqnames).

If ranges is not supplied and/or NULL then the constructor proceeds in 2 steps:

1. An initial GRanges object is created with as(seqnames, "GRanges").
2. Then this GRanges object is updated according to whatever non-NULL remaining arguments were passed to the call to GRanges().

Because of this behavior, GRanges(x) is equivalent to as(x, "GRanges").

Coercion

In the code snippets below, x is a GRanges object.

as(from, "GRanges"): Creates a GRanges object from a character vector, a factor, or a Ranged-Data, RangesList, RleList, or RleViewsList object.

When from is a vector (or factor), each element in it must represent a genomic range in format chr1:2501-2800 (unstranded range) or chr1:2501-2800:+ (stranded range). . . is also supported as a separator between the start and end positions. Strand can be +, -, *, or missing. The names on from are propagated to the returned GRanges object. See as.character() and as.factor() below for the reverse transformations.

Coercing a data.frame or DataFrame into a GRanges object is also supported. See [makeGRangesFromDataFrame](#) for the details.

- `as(from, "RangedData")`: Creates a RangedData object from a GRanges object. The strand and metadata columns become columns in the result. The `seqlengths(from)`, `isCircular(from)`, and `genome(from)` vectors are stored in the metadata columns of `ranges(rd)`.
- `as(from, "RangesList")`: Creates a RangesList object from a GRanges object. The strand and metadata columns become *inner* metadata columns (i.e. metadata columns on the ranges). The `seqlengths(from)`, `isCircular(from)`, and `genome(from)` vectors become the metadata columns.
- `as.character(x, ignore.strand=FALSE)`: Turn GRanges object `x` into a character vector where each range in `x` is represented by a string in format `chr1:2501-2800:+`. If `ignore.strand` is TRUE or if *all* the ranges in `x` are unstranded (i.e. their strand is set to `*`), then all the strings in the output are in format `chr1:2501-2800`.
The names on `x` are propagated to the returned character vector. Its metadata (`metadata(x)`) and metadata columns (`mcols(x)`) are ignored.
See `as(from, "GRanges")` above for the reverse transformation.
- `as.factor(x)`: Equivalent to
`factor(as.character(x), levels=as.character(sort(unique(x))))`
 See `as(from, "GRanges")` above for the reverse transformation.
 Note that `table(x)` is supported on a GRanges object. It is equivalent to, but much faster than, `table(as.factor(x))`.
- `as.data.frame(x, row.names = NULL, optional = FALSE, ...)`: Creates a data.frame with columns `seqnames` (factor), `start` (integer), `end` (integer), `width` (integer), `strand` (factor), as well as the additional metadata columns stored in `mcols(x)`. Pass an explicit `stringsAsFactors=TRUE/FALSE` argument via `...` to override the default conversions for the metadata columns in `mcols(x)`.

In the code snippets below, `x` is a [Seqinfo](#) object.

- `as(x, "GRanges")`, `as(x, "GenomicRanges")`, `as(x, "RangesList")`: Turns [Seqinfo](#) object `x` (with no NA lengths) into a GRanges or RangesList.

Accessors

In the following code snippets, `x` is a GRanges object.

- `length(x)`: Get the number of elements.
- `seqnames(x)`, `seqnames(x) <- value`: Get or set the sequence names. `value` can be an [Rle](#) object, a character vector, or a factor.
- `ranges(x)`, `ranges(x) <- value`: Get or set the ranges. `value` can be a Ranges object.
- `names(x)`, `names(x) <- value`: Get or set the names of the elements.
- `strand(x)`, `strand(x) <- value`: Get or set the strand. `value` can be an [Rle](#) object, character vector, or factor.
- `mcols(x, use.names=FALSE)`, `mcols(x) <- value`: Get or set the metadata columns. If `use.names=TRUE` and the metadata columns are not NULL, then the names of `x` are propagated as the row names of the returned [DataFrame](#) object. When setting the metadata columns, the supplied value must be NULL or a data.frame-like object (i.e. [DataTable](#) or `data.frame`) object holding element-wise metadata.

`elementMetadata(x)`, `elementMetadata(x) <- value`, `values(x)`, `values(x) <- value`: Alternatives to `mcols` functions. Their use is discouraged.

`seqinfo(x)`, `seqinfo(x) <- value`: Get or set the information about the underlying sequences. `value` must be a [Seqinfo](#) object.

`seqlevels(x)`, `seqlevels(x, force=FALSE) <- value`: Get or set the sequence levels. `seqlevels(x)` is equivalent to `seqlevels(seqinfo(x))` or to `levels(seqnames(x))`, those 2 expressions being guaranteed to return identical character vectors on a `GRanges` object. `value` must be a character vector with no NAs. See [?seqlevels](#) for more information.

`seqlengths(x)`, `seqlengths(x) <- value`: Get or set the sequence lengths. `seqlengths(x)` is equivalent to `seqlengths(seqinfo(x))`. `value` can be a named non-negative integer or numeric vector eventually with NAs.

`isCircular(x)`, `isCircular(x) <- value`: Get or set the circularity flags. `isCircular(x)` is equivalent to `isCircular(seqinfo(x))`. `value` must be a named logical vector eventually with NAs.

`genome(x)`, `genome(x) <- value`: Get or set the genome identifier or assembly name for each sequence. `genome(x)` is equivalent to `genome(seqinfo(x))`. `value` must be a named character vector eventually with NAs.

`seqlevelsStyle(x)`, `seqlevelsStyle(x) <- value`: Get or set the seqname style for `x`. See the [seqlevelsStyle](#) generic getter and setter in the **GenomeInfoDb** package for more information.

`score(x)`, `score(x) <- value`: Get or set the “score” column from the element metadata.

`granges(x, use.mcols=FALSE)`: Gets a `GRanges` with only the range information from `x`, unless `use.mcols` is `TRUE`, in which case the metadata columns are also returned. Those columns will include any “extra column slots” if `x` is a specialized `GenomicRanges` derivative.

Ranges methods

In the following code snippets, `x` is a `GRanges` object.

`start(x)`, `start(x) <- value`: Get or set `start(ranges(x))`.

`end(x)`, `end(x) <- value`: Get or set `end(ranges(x))`.

`width(x)`, `width(x) <- value`: Get or set `width(ranges(x))`.

Splitting and Combining

In the code snippets below, `x` is a `GRanges` object.

`append(x, values, after = length(x))`: Inserts the `values` into `x` at the position given by `after`, where `x` and `values` are of the same class.

`c(x, ...)`: Combines `x` and the `GRanges` objects in `...` together. Any object in `...` must belong to the same class as `x`, or to one of its subclasses, or must be `NULL`. The result is an object of the same class as `x`.

`c(x, ..., ignore.mcols=FALSE)` If the `GRanges` objects have metadata columns (represented as one [DataFrame](#) per object), each such [DataFrame](#) must have the same columns in order to combine successfully. In order to circumvent this restraint, you can pass in an `ignore.mcols=TRUE` argument which will combine all the objects into one and drop all of their metadata columns.

`split(x, f, drop=FALSE)`: Splits `x` according to `f` to create a `GRangesList` object. If `f` is a list-like object then `drop` is ignored and `f` is treated as if it was `rep(seq_len(length(f)), sapply(f, length))`, so the returned object has the same shape as `f` (it also receives the names of `f`). Otherwise, if `f` is not a list-like object, empty list elements are removed from the returned object if `drop` is `TRUE`.

Subsetting

In the code snippets below, `x` is a `GRanges` object.

`x[i, j], x[i, j] <- value`: Get or set elements `i` with optional metadata columns `mcols(x)[, j]`, where `i` can be missing; an NA-free logical, numeric, or character vector; or a 'logical' Rle object.

`x[i, j] <- value`: Replaces elements `i` and optional metadata columns `j` with `value`.

`head(x, n = 6L)`: If `n` is non-negative, returns the first `n` elements of the `GRanges` object. If `n` is negative, returns all but the last `abs(n)` elements of the `GRanges` object.

`rep(x, times, length.out, each)`: Repeats the values in `x` through one of the following conventions:

`times` Vector giving the number of times to repeat each element if of length `length(x)`, or to repeat the whole vector if of length 1.

`length.out` Non-negative integer. The desired length of the output vector.

`each` Non-negative integer. Each element of `x` is repeated `each` times.

`subset(x, subset)`: Returns a new object of the same class as `x` made of the subset using logical vector `subset`, where missing values are taken as `FALSE`.

`tail(x, n = 6L)`: If `n` is non-negative, returns the last `n` elements of the `GRanges` object. If `n` is negative, returns all but the first `abs(n)` elements of the `GRanges` object.

`window(x, start = NA, end = NA, width = NA, frequency = NULL, delta = NULL, ...)`: Extracts the subsequence window from the `GRanges` object using:

`start, end, width` The start, end, or width of the window. Two of the three are required.

`frequency, delta` Optional arguments that specify the sampling frequency and increment within the window.

In general, this is more efficient than using `"["` operator.

`window(x, start = NA, end = NA, width = NA, keepLength = TRUE) <- value`: Replaces the subsequence window specified on the left (i.e. the subsequence in `x` specified by `start`, `end` and `width`) by `value`. `value` must either be of class `class(x)`, belong to a subclass of `class(x)`, be coercible to `class(x)`, or be `NULL`. If `keepLength` is `TRUE`, the elements of `value` are repeated to create a `GRanges` object with the same number of elements as the width of the subsequence window it is replacing. If `keepLength` is `FALSE`, this replacement method can modify the length of `x`, depending on how the length of the left subsequence window compares to the length of `value`.

`x$name, x$name <- value`: Shortcuts for `mcols(x)$name` and `mcols(x)$name <- value`, respectively. Provided as a convenience, for `GRanges` objects *only*, and as the result of strong popular demand. Note that those methods are not consistent with the other `$` and `$<-` methods in the `IRanges/GenomicRanges` infrastructure, and might confuse some users by making them believe that a `GRanges` object can be manipulated as a data.frame-like object. Therefore we

recommend using them only interactively, and we discourage their use in scripts or packages. For the latter, use `mcols(x)$name` and `mcols(x)$name <- value`, instead of `x$name` and `x$name <- value`, respectively.

Note that a GRanges object can be used to as a subscript to subset a list-like object that has names on it. In that case, the names on the list-like object are interpreted as sequence names. In the code snippets below, `x` is a list or [List](#) object with names on it, and the subscript `gr` is a GRanges object with all its seqnames being valid `x` names.

`x[gr]`: Return an object of the same class as `x` and *parallel* to `gr`. More precisely, it's conceptually doing:

```
lapply(gr, function(gr1) x[[seqnames(gr1)]] [ranges(gr1)])
```

Other methods

`show(x)`: By default the `show` method displays 5 head and 5 tail elements. This can be changed by setting the global options `showHeadLines` and `showTailLines`. If the object length is less than (or equal to) the sum of these 2 options plus 1, then the full object is displayed. Note that these options also affect the display of [GAlignments](#) and [GAlignmentPairs](#) objects (defined in the **GenomicAlignments** package), as well as other objects defined in the **IRanges** and **Biostrings** packages (e.g. [IRanges](#) and [DNAStringSet](#) objects).

Author(s)

P. Aboyoun and H. Pages

See Also

- [makeGRangesFromDataFrame](#)
- [seqinfo](#)
- [GenomicRanges-comparison](#)
- [intra-range-methods](#)
- [inter-range-methods](#)
- [coverage-methods](#)
- [setops-methods](#)
- [findOverlaps-methods](#)
- [nearest-methods](#)
- [absoluteRanges](#)
- [tileGenome](#)
- [GRangesList-class](#)
- [Ranges-class](#)
- [Vector-class](#)
- [Rle-class](#)
- [DataFrame-class](#)

Examples

```
## -----
## CONSTRUCTION
## -----
## Specifying the bare minimum i.e. seqnames and ranges only. The
## GRanges object will have no names, no strand information, and no
## metadata columns:
gr0 <- GRanges(Rle(c("chr2", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
               IRanges(1:10, width=10:1))

gr0

## Specifying names, strand, metadata columns. They can be set on an
## existing object:
names(gr0) <- head(letters, 10)
strand(gr0) <- Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2))
mcols(gr0)$score <- 1:10
mcols(gr0)$GC <- seq(1, 0, length=10)
gr0

## ... or specified at construction time:
gr <- GRanges(Rle(c("chr2", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
              IRanges(1:10, width=10:1, names=head(letters, 10)),
              Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
              score=1:10, GC=seq(1, 0, length=10))
stopifnot(identical(gr0, gr))

## Specifying the seqinfo. It can be set on an existing object:
seqinfo <- Seqinfo(paste0("chr", 1:3), c(1000, 2000, 1500), NA, "mock1")
seqinfo(gr0) <- merge(seqinfo(gr0), seqinfo)
seqlevels(gr0) <- seqlevels(seqinfo)

## ... or specified at construction time:
gr <- GRanges(Rle(c("chr2", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
              IRanges(1:10, width=10:1, names=head(letters, 10)),
              Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
              score=1:10, GC=seq(1, 0, length=10),
              seqinfo=seqinfo)
stopifnot(identical(gr0, gr))

## -----
## COERCION
## -----
## From GRanges:
as.character(gr)
as.factor(gr)
as.data.frame(gr)

## From character to GRanges:
x1 <- "chr2:56-125"
as(x1, "GRanges")
as(rep(x1, 4), "GRanges")
x2 <- c(A=x1, B="chr1:25-30:-")
```



```

as(x2, "GRanges")

## From data.frame to GRanges:
df <- data.frame(chrom="chr2", start=11:15, end=20:24)
gr3 <- as(df, "GRanges")

## Alternatively, coercion to GRanges can be done by just calling the
## GRanges() constructor on the object to coerce:
gr1 <- GRanges(x1) # same as as(x1, "GRanges")
gr2 <- GRanges(x2) # same as as(x2, "GRanges")
gr3 <- GRanges(df) # same as as(df, "GRanges")

## Sanity checks:
stopifnot(identical(as(x1, "GRanges"), gr1))
stopifnot(identical(as(x2, "GRanges"), gr2))
stopifnot(identical(as(df, "GRanges"), gr3))

## -----
## SUMMARIZING ELEMENTS
## -----
table(seqnames(gr))
table(strand(gr))
sum(width(gr))
table(gr)
summary(mcols(gr)[,"score"])

## The number of lines displayed in the 'show' method are controlled
## with two global options:
longGR <- sample(gr, 25, replace=TRUE)
longGR
options(showHeadLines=7)
options(showTailLines=2)
longGR

## Revert to default values
options(showHeadLines=NULL)
options(showTailLines=NULL)

## -----
## RENAMING THE UNDERLYING SEQUENCES
## -----
seqlevels(gr)
seqlevels(gr) <- sub("chr", "Chrom", seqlevels(gr))
gr
seqlevels(gr) <- sub("Chrom", "chr", seqlevels(gr)) # revert

## -----
## COMBINING OBJECTS
## -----
gr2 <- GRanges(seqnames=Rle(c('chr1', 'chr2', 'chr3'), c(3, 3, 4)),
               IRanges(1:10, width=5),
               strand="-",
               score=101:110, GC=runif(10),

```

```

      seqinfo=seqinfo)
gr3 <- GRanges(seqnames=Rle(c('chr1', 'chr2', 'chr3'), c(3, 4, 3)),
              IRanges(101:110, width=10),
              strand='-',
              score=21:30,
              seqinfo=seqinfo)
some.gr <- c(gr, gr2)

## all.gr <- c(gr, gr2, gr3) ## (This would fail)
all.gr <- c(gr, gr2, gr3, ignore.mcols=TRUE)

## -----
## USING A GRANGES OBJECT AS A SUBSCRIPT TO SUBSET ANOTHER OBJECT
## -----
## Subsetting *by* a GRanges subscript is supported only if the object
## to subset is a named list-like object:
x <- RleList(chr1=101:120, chr2=2:-8, chr3=31:40)
x[gr]

```

GRangesList-class

GRangesList objects

Description

The GRangesList class is a container for storing a collection of GRanges objects. It is derived from GenomicRangesList.

Constructors

`GRangesList(...)`: Creates a GRangesList object using GRanges objects supplied in

`makeGRangesListFromFeatureFragments(seqnames=Rle(factor()), fragmentStarts=list(), fragmentEnds=list(), ...)`
Constructs a GRangesList object from a list of fragmented features. See the Examples section below.

Accessors

In the following code snippets, `x` is a GRanges object.

`length(x)`: Get the number of list elements.

`names(x), names(x) <- value`: Get or set the names on `x`.

`elementLengths(x)`: Get the length of each of the list elements.

`isEmpty(x)`: Returns a logical indicating either if the GRangesList has no elements or if all its elements are empty.

`seqnames(x), seqnames(x) <- value`: Get or set the sequence names in the form of an RleList. `value` can be an RleList or CharacterList object.

`ranges(x, use.mcols=FALSE), ranges(x) <- value`: Get or set the ranges in the form of a CompressedIRangesList. `value` can be a RangesList object.

`start(x)`, `start(x) <- value`: Get or set `start(ranges(x))`.
`end(x)`, `end(x) <- value`: Get or set `end(ranges(x))`.
`width(x)`, `width(x) <- value`: Get or set `width(ranges(x))`.
`strand(x)`, `strand(x) <- value`: Get or set the strand in the form of an `RleList`. `value` can be an `RleList`, `CharacterList` or single character. `value` as a single character converts all ranges in `x` to the same value; for selective strand conversion (i.e., mixed “+” and “-”) use `RleList` or `CharacterList`.
`mcols(x, use.names=FALSE)`, `mcols(x) <- value`: Get or set the metadata columns. `value` can be `NULL`, or a data.frame-like object (i.e. [DataFrame](#) or `data.frame`) holding element-wise metadata.
`elementMetadata(x)`, `elementMetadata(x) <- value`, `values(x)`, `values(x) <- value`: Alternatives to `mcols` functions. Their use is discouraged.
`seqinfo(x)`, `seqinfo(x) <- value`: Get or set the information about the underlying sequences. `value` must be a [Seqinfo](#) object.
`seqlevels(x)`, `seqlevels(x, force=FALSE) <- value`: Get or set the sequence levels. `seqlevels(x)` is equivalent to `seqlevels(seqinfo(x))` or to `levels(seqnames(x))`, those 2 expressions being guaranteed to return identical character vectors on a `GRangesList` object. `value` must be a character vector with no NAs. See [?seqlevels](#) for more information.
`seqlengths(x)`, `seqlengths(x) <- value`: Get or set the sequence lengths. `seqlengths(x)` is equivalent to `seqlengths(seqinfo(x))`. `value` can be a named non-negative integer or numeric vector eventually with NAs.
`isCircular(x)`, `isCircular(x) <- value`: Get or set the circularity flags. `isCircular(x)` is equivalent to `isCircular(seqinfo(x))`. `value` must be a named logical vector eventually with NAs.
`genome(x)`, `genome(x) <- value`: Get or set the genome identifier or assembly name for each sequence. `genome(x)` is equivalent to `genome(seqinfo(x))`. `value` must be a named character vector eventually with NAs.
`seqlevelsStyle(x)`, `seqlevelsStyle(x) <- value`: Get or set the seqname style for `x`. See the [seqlevelsStyle](#) generic getter and setter in the **GenomeInfoDb** package for more information.
`score(x)`, `score(x) <- value`: Get or set the “score” metadata column.

Coercion

In the code snippets below, `x` is a `GRangesList` object.

`as.data.frame(x, row.names = NULL, optional = FALSE, ..., value.name = "value", use.outer.mcols = FALSE)`
 Coerces `x` to a data.frame. See `as.data.frame` on the `List` man page for details (`?List`).
`as.list(x, use.names = TRUE)`: Creates a list containing the elements of `x`.
`as(x, "IRangesList")`: Turns `x` into an [IRangesList](#) object.
`as(from, "GRangesList")`: Creates a `GRangesList` object from a [RangedDataList](#) object.

Subsetting

In the following code snippets, `x` is a `GRangesList` object.

`x[i, j], x[i, j] <- value`: Get or set elements `i` with optional metadata columns `mcols(x)[, j]`, where `i` can be missing; an NA-free logical, numeric, or character vector; a 'logical' Rle object, or an `AtomicList` object.

`x[[i]], x[[i]] <- value`: Get or set element `i`, where `i` is a numeric or character vector of length 1.

`x$name, x$name <- value`: Get or set element name, where `name` is a name or character vector of length 1.

`head(x, n = 6L)`: If `n` is non-negative, returns the first `n` elements of the `GRangesList` object. If `n` is negative, returns all but the last `abs(n)` elements of the `GRangesList` object.

`rep(x, times, length.out, each)`: Repeats the values in `x` through one of the following conventions:

`times` Vector giving the number of times to repeat each element if of length `length(x)`, or to repeat the whole vector if of length 1.

`length.out` Non-negative integer. The desired length of the output vector.

`each` Non-negative integer. Each element of `x` is repeated `each` times.

`subset(x, subset)`: Returns a new object of the same class as `x` made of the subset using logical vector `subset`, where missing values are taken as `FALSE`.

`tail(x, n = 6L)`: If `n` is non-negative, returns the last `n` elements of the `GRanges` object. If `n` is negative, returns all but the first `abs(n)` elements of the `GRanges` object.

Combining

In the code snippets below, `x` is a `GRangesList` object.

`c(x, ...)`: Combines `x` and the `GRangesList` objects in `...` together. Any object in `...` must belong to the same class as `x`, or to one of its subclasses, or must be `NULL`. The result is an object of the same class as `x`.

`append(x, values, after = length(x))`: Inserts the `values` into `x` at the position given by `after`, where `x` and `values` are of the same class.

`unlist(x, recursive = TRUE, use.names = TRUE)`: Concatenates the elements of `x` into a single `GRanges` object.

Looping

In the code snippets below, `x` is a `GRangesList` object.

`endoapply(X, FUN, ...)`: Similar to `lapply`, but performs an endomorphism, i.e. returns an object of `class(X)`.

`lapply(X, FUN, ...)`: Like the standard `lapply` function defined in the base package, the `lapply` method for `GRangesList` objects returns a list of the same length as `X`, with each element being the result of applying `FUN` to the corresponding element of `X`.

`Map(f, ...)`: Applies a function to the corresponding elements of given `GRangesList` objects.

`mapply(FUN, ..., MoreArgs = NULL, SIMPLIFY = TRUE, USE.NAMES = TRUE)`: Like the standard `mapply` function defined in the base package, the `mapply` method for `GRangesList` objects is a multivariate version of `sapply`.

`mendoapply(FUN, ..., MoreArgs = NULL)`: Similar to `mapply`, but performs an endomorphism across multiple objects, i.e. returns an object of class `(list(...)[[1]])`.

`Reduce(f, x, init, right = FALSE, accumulate = FALSE)`: Uses a binary function to successively combine the elements of `x` and a possibly given initial value.

f A binary argument function.

init An R object of the same kind as the elements of `x`.

right A logical indicating whether to proceed from left to right (default) or from right to left.

nomatch The value to be returned in the case when "no match" (no element satisfying the predicate) is found.

`sapply(X, FUN, ..., simplify=TRUE, USE.NAMES=TRUE)`: Like the standard `sapply` function defined in the base package, the `sapply` method for `GRangesList` objects is a user-friendly version of `lapply` by default returning a vector or matrix if appropriate.

Author(s)

P. Aboyoun & H. Pages

See Also

[GRanges-class](#), [seqinfo](#), [Vector-class](#), [RangesList-class](#), [RleList-class](#), [DataFrameList-class](#), [intra-range-methods](#), [inter-range-methods](#), [coverage-methods](#), [setops-methods](#), [findOverlaps-methods](#)

Examples

```
## Construction with GRangesList():
gr1 <-
  GRanges(seqnames = "chr2", ranges = IRanges(3, 6),
          strand = "+", score = 5L, GC = 0.45)
gr2 <-
  GRanges(seqnames = c("chr1", "chr1"),
          ranges = IRanges(c(7,13), width = 3),
          strand = c("+", "-"), score = 3:4, GC = c(0.3, 0.5))
gr3 <-
  GRanges(seqnames = c("chr1", "chr2"),
          ranges = IRanges(c(1, 4), c(3, 9)),
          strand = c("-", "-"), score = c(6L, 2L), GC = c(0.4, 0.1))
grl <- GRangesList("gr1" = gr1, "gr2" = gr2, "gr3" = gr3)
grl

## Summarizing elements:
elementLengths(grl)
table(seqnames(grl))

## Extracting subsets:
grl[seqnames(grl) == "chr1", ]
grl[seqnames(grl) == "chr1" & strand(grl) == "+", ]
```

```

## Renaming the underlying sequences:
seqlevels(grl)
seqlevels(grl) <- sub("chr", "Chrom", seqlevels(grl))
grl

## Coerce to IRangesList (seqnames and strand information is lost):
as(grl, "IRangesList")

## isDisjoint():
isDisjoint(grl)

## disjoint():
disjoin(grl) # metadata columns and order NOT preserved

## Construction with makeGRangesListFromFeatureFragments():
filepath <- system.file("extdata", "feature_fragments.txt",
                        package="GenomicRanges")
featfrags <- read.table(filepath, header=TRUE, stringsAsFactors=FALSE)
grl2 <- with(featfrags,
            makeGRangesListFromFeatureFragments(seqnames=targetName,
                                                fragmentStarts=targetStart,
                                                fragmentWidths=blockSizes,
                                                strand=strand))

names(grl2) <- featfrags$RefSeqID
grl2

```

GRangesMapping-class *GRangesMapping objects*

Description

The `GRangesMapping` class is a container for both the genomic locations and the matching results output from the `map` function.

Details

The `map` generic converts a set of ranges to the equivalent ranges on another sequence, through some sort of alignment between sequences, and outputs a `GRangesMapping` object. There are two components of that object: the transformed `GRanges`, and the hits, a `Hits` object of the same length that matches each input range to a destination sequence (useful when the alignment is one/many to many).

GRangesMapping Accessors

`ranges(x)`: Gets the mapped `GRanges`.

`hits(x)`: Gets the matching between the input ranges and the destination sequences (of which there may be more than one).

`dim(x)`: Same as `dim(hits(x))`.

length(x): Same as length(hits(x)).
 subjectHits(x): Same as subjectHits(hits(x)).
 queryHits(x): Same as queryHits(hits(x)).

GRangesMapping **Coercion**

as(from, "RangedData"): Converts a GRangesMapping into a [RangedData](#). The ranges/space in the RangedData are the ranges/space of from, and the values result from the coercion of the hits to a DataFrame.
 as(from, "GenomicRanges"): Returns the [GRanges](#) with the hits (coerced to a DataFrame) as the mcols.

Author(s)

Michael Lawrence

See Also

Methods on the generic map, which generates an instance of this class, are defined in other packages, like [GenomicRanges](#).

Examples

```
# See ?map-methods in GenomicRanges or GenomicAlignments for examples.
```

inter-range-methods *Inter range transformations of a GenomicRanges or GRangesList object*

Description

See `?`intra-range-methods`` and `?`inter-range-methods`` in the [IRanges](#) package for a quick introduction to intra range and inter range transformations.

This man page documents inter range transformations of a [GenomicRanges](#) object (i.e. of an object that belongs to the [GenomicRanges](#) class or one of its subclasses, this includes for example [GRanges](#) objects).

See `?`intra-range-methods`` for intra range transformations of a [GenomicRanges](#) object.

Usage

```
## S4 method for signature 'GenomicRanges'
range(x, ..., ignore.strand=FALSE, na.rm=FALSE)
## S4 method for signature 'GRangesList'
range(x, ..., na.rm=FALSE)

## S4 method for signature 'GenomicRanges'
```

```

reduce(x, drop.empty.ranges=FALSE, min.gapwidth=1L,
       with.revmap=FALSE, with.inframe.attrib=FALSE,
       ignore.strand=FALSE)
## S4 method for signature 'GRangesList'
reduce(x, drop.empty.ranges=FALSE, min.gapwidth=1L,
       with.inframe.attrib=FALSE)

## S4 method for signature 'GenomicRanges'
gaps(x, start=1L, end=seqlengths(x))

## S4 method for signature 'GenomicRanges'
disjoin(x, ignore.strand=FALSE)
## S4 method for signature 'GRangesList'
disjoin(x, ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges'
isDisjoint(x, ignore.strand=FALSE)
## S4 method for signature 'GRangesList'
isDisjoint(x, ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges'
disjointBins(x, ignore.strand=FALSE)

```

Arguments

<code>x</code>	A GenomicRanges object.
<code>drop.empty.ranges</code> , <code>min.gapwidth</code> , <code>with.revmap</code> , <code>with.inframe.attrib</code> , <code>start</code> , <code>end</code>	See ?`inter-range-methods` in the <code>IRanges</code> package.
<code>ignore.strand</code>	TRUE or FALSE. Whether the strand of the input ranges should be ignored or not. See details below.
<code>...</code>	For range, additional <code>GenomicRanges</code> objects to consider. Ignored otherwise.
<code>na.rm</code>	Ignored.

Details

On a `GRanges` object: `range` returns an object of the same type as `x` containing range bounds for each distinct (seqname, strand) pairing. The names (`names(x)`) and the metadata columns in `x` are dropped.

`reduce` returns an object of the same type as `x` containing reduced ranges for each distinct (seqname, strand) pairing. The names (`names(x)`) and the metadata columns in `x` are dropped. See [?reduce](#) for more information about range reduction and for a description of the optional arguments.

`gaps` returns an object of the same type as `x` containing complemented ranges for each distinct (seqname, strand) pairing. The names (`names(x)`) and the columns in `x` are dropped. For the start and end arguments of this `gaps` method, it is expected that the user will supply a named integer vector (where the names correspond to the appropriate seqlevels). See [?gaps](#) for more information about range complements and for a description of the optional arguments.

`disjoin` returns an object of the same type as `x` containing disjoint ranges for each distinct (seqname, strand) pairing. The names (`names(x)`) and the metadata columns in `x` are dropped.

`isDisjoint` returns a logical value indicating whether the ranges in `x` are disjoint (i.e. non-overlapping).

`disjointBins` returns bin indexes for the ranges in `x`, such that ranges in the same bin do not overlap. If `ignore.strand=FALSE`, the two features cannot overlap if they are on different strands.

On a `GRangesList` object: When they are supported on `GRangesList` object `x`, the above inter range transformations will apply the transformation to each of the list elements in `x` and return a list-like object *parallel* to `x` (i.e. with 1 list element per list element in `x`). If `x` has names on it, they're propagated to the returned object.

Author(s)

H. Pages and P. Aboyoun

See Also

- The [GenomicRanges](#) and [GRanges](#) classes.
- The [Ranges](#) class in the `IRanges` package.
- The [inter-range-methods](#) man page in the `IRanges` package.
- [GenomicRanges-comparison](#) for comparing and ordering genomic ranges.

Examples

```
gr <- GRanges(
  seqnames=Rle(paste("chr", c(1, 2, 1, 3), sep=""), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, width=10:1, names=letters[1:10]),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  score=1:10,
  GC=seq(1, 0, length=10)
)
gr

gr1 <- GRanges(seqnames="chr2", ranges=IRanges(3, 6),
  strand="+", score=5L, GC=0.45)
gr2 <- GRanges(seqnames=c("chr1", "chr1"),
  ranges=IRanges(c(7,13), width=3),
  strand=c("+", "-"), score=3:4, GC=c(0.3, 0.5))
gr3 <- GRanges(seqnames=c("chr1", "chr2"),
  ranges=IRanges(c(1, 4), c(3, 9)),
  strand=c("-", "-"), score=c(6L, 2L), GC=c(0.4, 0.1))
grl <- GRangesList(gr1=gr1, gr2=gr2, gr3=gr3)
grl

## -----
## range()
## -----

## On a GRanges object:
```

```

range(gr)

## On a GRangesList object:
range(gr1)

# -----
## reduce()
## -----
reduce(gr)

gr2 <- reduce(gr, with.revmap=TRUE)
revmap <- mcols(gr2)$revmap # an IntegerList

## Use the mapping from reduced to original ranges to group the original
## ranges by reduced range:
relist(gr[unlist(revmap)], revmap)

## Or use it to split the DataFrame of original metadata columns by
## reduced range:
relist(mcols(gr)[unlist(revmap), ], revmap) # a SplitDataFrameList

## [For advanced users] Use this reverse mapping to compare the reduced
## ranges with the ranges they originate from:
expanded_gr2 <- rep(gr2, elementLengths(revmap))
reordered_gr <- gr[unlist(revmap)]
codes <- compare(expanded_gr2, reordered_gr)
## All the codes should translate to "d", "e", "g", or "h" (the 4 letters
## indicating that the range on the left contains the range on the right):
alphacodes <- rangeComparisonCodeToLetter(compare(expanded_gr2, reordered_gr))
stopifnot(all(alphacodes %in% c("d", "e", "g", "h")))

## On a big GRanges object with a lot of seqlevels:
mcols(gr) <- NULL
biggr <- c(gr, GRanges("chr1", IRanges(c(4, 1), c(5, 2)), strand="+"))
seqlevels(biggr) <- paste0("chr", 1:2000)
biggr <- rep(biggr, 25000)
set.seed(33)
seqnames(biggr) <- sample(factor(seqlevels(biggr), levels=seqlevels(biggr)),
                           length(biggr), replace=TRUE)

biggr2 <- reduce(biggr, with.revmap=TRUE)
revmap <- mcols(biggr2)$revmap
expanded_biggr2 <- rep(biggr2, elementLengths(revmap))
reordered_biggr <- biggr[unlist(revmap)]
codes <- compare(expanded_biggr2, reordered_biggr)
alphacodes <- rangeComparisonCodeToLetter(compare(expanded_biggr2,
                                                reordered_biggr))
stopifnot(all(alphacodes %in% c("d", "e", "g", "h")))
table(alphacodes)

## On a GRangesList object:
reduce(gr1) # Doesn't really reduce anything but note the reordering
            # of the inner elements in the 3rd top-level element: the

```

```

# ranges are reordered by sequence name first (the order of
# the sequence names is dictated by the sequence levels),
# and then by strand.

## -----
## gaps()
## -----
gaps(gr, start=1, end=10)

## -----
## disjoint(), isDisjoint(), disjointBins()
## -----
disjoin(gr)
isDisjoint(gr)
stopifnot(isDisjoint(disjoin(gr)))
disjointBins(gr)
stopifnot(all(sapply(split(gr, disjointBins(gr)), isDisjoint)))

```

intra-range-methods *Intra range transformations of a GRanges or GRangesList object*

Description

This man page documents intra range transformations of a [GenomicRanges](#) object (i.e. of an object that belongs to the [GenomicRanges](#) class or one of its subclasses, this includes for example [GRanges](#) objects), or a [GRangesList](#) object.

See `?`intra-range-methods`` and `?`inter-range-methods`` in the **IRanges** package for a quick introduction to intra range and inter range transformations.

Intra range methods for [GAlignments](#) and [GAlignmentsList](#) objects are defined and documented in the **GenomicAlignments** package.

See `?`inter-range-methods`` for inter range transformations of a [GenomicRanges](#) or [GRangesList](#) object.

Usage

```

## S4 method for signature 'GenomicRanges'
shift(x, shift=0L, use.names=TRUE)
## S4 method for signature 'GRangesList'
shift(x, shift=0L, use.names=TRUE)

## S4 method for signature 'GenomicRanges'
narrow(x, start=NA, end=NA, width=NA, use.names=TRUE)

## S4 method for signature 'GenomicRanges'
resize(x, width, fix="start", use.names=TRUE,
       ignore.strand=FALSE)
## S4 method for signature 'GRangesList'

```

```

resize(x, width, fix="start", use.names=TRUE,
       ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges'
flank(x, width, start=TRUE, both=FALSE,
      use.names=TRUE, ignore.strand=FALSE)
## S4 method for signature 'GRangesList'
flank(x, width, start=TRUE, both=FALSE,
      use.names=TRUE, ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges'
promoters(x, upstream=2000, downstream=200, ...)
## S4 method for signature 'GRangesList'
promoters(x, upstream=2000, downstream=200, ...)

## S4 method for signature 'GenomicRanges'
restrict(x, start=NA, end=NA, keep.all.ranges=FALSE,
        use.names=TRUE)
## S4 method for signature 'GRangesList'
restrict(x, start=NA, end=NA, keep.all.ranges=FALSE,
        use.names=TRUE)

## S4 method for signature 'GenomicRanges'
trim(x, use.names=TRUE)

```

Arguments

`x` A [GenomicRanges](#) or [GRangesList](#) object.

`shift`, `use.names`, `start`, `end`, `width`, `both`, `fix`, `keep.all.ranges`, `upstream`, `downstream`
See ?`intra-range-methods`.

`ignore.strand` TRUE or FALSE. Whether the strand of the input ranges should be ignored or not.
See details below.

... Additional arguments to methods.

Details

- `shift` behaves like the `shift` method for [Ranges](#) objects. See ?`intra-range-methods` for the details.
- `()narrow` on a [GenomicRanges](#) object behaves like on a [Ranges](#) object. See ?`intra-range-methods` for the details.
A major difference though is that it returns a [GenomicRanges](#) object instead of a [Ranges](#) object. The returned object is *parallel* (i.e. same length and names) to the original object `x`.
- `resize` returns an object of the same type and length as `x` containing intervals that have been resized to width `width` based on the `strand(x)` values. Elements where `strand(x) == "+"` or `strand(x) == "*"` are anchored at `start(x)` and elements where `strand(x) == "-"` are anchored at the `end(x)`. The `use.names` argument determines whether or not to keep the names on the ranges.

- `flank` returns an object of the same type and length as `x` containing intervals of width `width` that flank the intervals in `x`. The `start` argument takes a logical indicating whether `x` should be flanked at the "start" (TRUE) or the "end" (FALSE), which for `strand(x) != "-"` is `start(x)` and `end(x)` respectively and for `strand(x) == "-"` is `end(x)` and `start(x)` respectively. The `both` argument takes a single logical value indicating whether the flanking region width positions extends *into* the range. If `both=TRUE`, the resulting range thus straddles the end point, with `width` positions on either side.
- `promoters` returns an object of the same type and length as `x` containing promoter ranges. Promoter ranges extend around the transcription start site (TSS) which is defined as `start(x)`. The `upstream` and `downstream` arguments define the number of nucleotides in the 5' and 3' direction, respectively. The full range is defined as, $(start(x) - upstream)$ to $(start(x) + downstream - 1)$.
Ranges on the `*` strand are treated the same as those on the `+` strand. When no `seqlengths` are present in `x`, it is possible to have non-positive start values in the promoter ranges. This occurs when $(TSS - upstream) < 1$. In the equal but opposite case, the end values of the ranges may extend beyond the chromosome end when $(TSS + downstream + 1) > \text{'chromosome end'}$. When `seqlengths` are not `NA` the promoter ranges are kept within the bounds of the defined `seqlengths`.
- `restrict` returns an object of the same type and length as `x` containing restricted ranges for distinct `seqnames`. The `start` and `end` arguments can be a named numeric vector of `seqnames` for the ranges to be restricted or a numeric vector of length 1 if the restriction operation is to be applied to all the sequences in `x`. See `?`intra-range-methods`` for more information about range restriction and for a description of the optional arguments.
- `trim` trims out-of-bound ranges located on non-circular sequences whose length is not `NA`.

Author(s)

P. Aboyoun and V. Obenchain <vobencha@fhcrc.org>

See Also

- [GenomicRanges](#), [GRanges](#), and [GRangesList](#) objects.
- The [intra-range-methods](#) man page in the **IRanges** package.
- The [Ranges](#) class in the **IRanges** package.

Examples

```
## -----
## A. ON A GRanges OBJECT
## -----
gr <- GRanges(
  seqnames=Rle(paste("chr", c(1, 2, 1, 3), sep=""), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, width=10:1, names=letters[1:10]),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  score=1:10,
  GC=seq(1, 0, length=10)
)
gr
```


Arguments

<code>df</code>	A data.frame or DataFrame object. If not, then the function tries to turn <code>df</code> into a data frame with <code>as.data.frame(df)</code> .
<code>keep.extra.columns</code>	TRUE or FALSE (the default). If TRUE, the columns in <code>df</code> that are not used to form the genomic ranges of the returned GRanges object are then returned as metadata columns on the object. Otherwise, they are ignored. If <code>df</code> has a width column, then it's always ignored.
<code>ignore.strand</code>	TRUE or FALSE (the default). If TRUE, then the strand of the returned GRanges object is set to "*".
<code>seqinfo</code>	Either NULL, or a Seqinfo object, or a character vector of seqlevels, or a named numeric vector of sequence lengths. When not NULL, it must be compatible with the genomic ranges in <code>df</code> i.e. it must include at least the sequence levels represented in <code>df</code> .
<code>seqnames.field</code>	A character vector of recognized names for the column in <code>df</code> that contains the chromosome name (a.k.a. sequence name) associated with each genomic range. Only the first name in <code>seqnames.field</code> that is found in <code>colnames(df)</code> is used. If no one is found, then an error is raised.
<code>start.field</code>	A character vector of recognized names for the column in <code>df</code> that contains the start positions of the genomic ranges. Only the first name in <code>start.field</code> that is found in <code>colnames(df)</code> is used. If no one is found, then an error is raised.
<code>end.field</code>	A character vector of recognized names for the column in <code>df</code> that contains the end positions of the genomic ranges. Only the first name in <code>start.field</code> that is found in <code>colnames(df)</code> is used. If no one is found, then an error is raised.
<code>strand.field</code>	A character vector of recognized names for the column in <code>df</code> that contains the strand associated with each genomic range. Only the first name in <code>strand.field</code> that is found in <code>colnames(df)</code> is used. If no one is found or if <code>ignore.strand</code> is TRUE, then the strand of the returned GRanges object is set to "*".
<code>starts.in.df.are.0based</code>	TRUE or FALSE (the default). If TRUE, then the start positions of the genomic ranges in <code>df</code> are considered to be <i>0-based</i> and are converted to <i>1-based</i> in the returned GRanges object. This feature is intended to make it more convenient to handle input that contains data obtained from resources using the "0-based start" convention. A notorious example of such resource is the UCSC Table Browser (http://genome.ucsc.edu/cgi-bin/hgTables).

Value

A [GRanges](#) object with one element per row in the input.

If the `seqinfo` argument was supplied, the returned object will have exactly the seqlevels specified in `seqinfo` and in the same order. Otherwise, the seqlevels are ordered according to the output of the [rankSeqlevels](#) function (except if `df` contains the `seqnames` in the form of a factor-Rle, in which case the levels of the factor-Rle become the seqlevels of the returned object and with no re-ordering).

If `df` has non-automatic row names (i.e. `rownames(df)` is not NULL and is not `seq_len(nrow(df))`), then they will be used to set names on the returned [GRanges](#) object.

Note

Coercing `data.frame` or `DataFrame` `df` into a `GRanges` object (with `as(df, "GRanges")`) is equivalent to calling `makeGRangesFromDataFrame(df, keep.extra.columns=TRUE)`.

Author(s)

H. Pages, based on a proposal by Kasper Daniel Hansen

See Also

- `GRanges` objects.
- `Seqinfo` objects and the `rankSeqlevels` function in the **GenomeInfoDb** package.
- The `makeGRangesListFromFeatureFragments` function for making a `GRangesList` object from a list of fragmented features.
- The `getTable` function in the **rtracklayer** package for an R interface to the UCSC Table Browser.
- `DataFrame` objects in the **S4Vectors** package.

Examples

```
## -----
## BASIC EXAMPLES
## -----

df <- data.frame(chr="chr1", start=11:15, end=12:16,
                 strand=c("+", "-", "+", "*", "."), score=1:5)
df
makeGRangesFromDataFrame(df) # strand value "." is replaced with "*"

## The strand column is optional:
df <- data.frame(chr="chr1", start=11:15, end=12:16, score=1:5)
makeGRangesFromDataFrame(df)

gr <- makeGRangesFromDataFrame(df, keep.extra.columns=TRUE)
gr2 <- as(df, "GRanges") # equivalent to the above
stopifnot(identical(gr, gr2))

makeGRangesFromDataFrame(df, ignore.strand=TRUE)
makeGRangesFromDataFrame(df, keep.extra.columns=TRUE,
                          ignore.strand=TRUE)

makeGRangesFromDataFrame(df, seqinfo=paste0("chr", 4:1))
makeGRangesFromDataFrame(df, seqinfo=c(chrM=NA, chr1=500, chrX=100))
makeGRangesFromDataFrame(df, seqinfo=Seqinfo(paste0("chr", 4:1)))

## -----
## ABOUT AUTOMATIC DETECTION OF THE seqnames/start/end/strand COLUMNS
## -----

## Automatic detection of the seqnames/start/end/strand columns is
```



```

## case insensitive:
df <- data.frame(ChROM="chr1", Start=11:15, stop=12:16,
                 STRAND=c("+","-","+","*","."), score=1:5)
makeGRangesFromDataFrame(df)

## It also ignores a common prefix between the start and end columns:
df <- data.frame(seqnames="chr1", tx_start=11:15, tx_end=12:16,
                 strand=c("+","-","+","*","."), score=1:5)
makeGRangesFromDataFrame(df)

## The common prefix between the start and end columns is used to
## disambiguate between more than one seqnames column:
df <- data.frame(chrom="chr1", tx_start=11:15, tx_end=12:16,
                 tx_chr="chr2", score=1:5)
makeGRangesFromDataFrame(df)

## -----
## 0-BASED VS 1-BASED START POSITIONS
## -----

if (require(rtracklayer)) {
  session <- browserSession()
  genome(session) <- "sacCer2"
  query <- ucscTableQuery(session, "Most Conserved")
  df <- getTable(query)

  ## A common pitfall is to forget that the UCSC Table Browser uses the
  ## "0-based start" convention:
  gr0 <- makeGRangesFromDataFrame(df, keep.extra.columns=TRUE)
  head(gr0)
  min(start(gr0))

  ## The start positions need to be converted into 1-based positions,
  ## to adhere to the convention used in Bioconductor:
  gr1 <- makeGRangesFromDataFrame(df, keep.extra.columns=TRUE,
                                 starts.in.df.are.0based=TRUE)

  head(gr1)
}

```

```
makeSummarizedExperimentFromExpressionSet
```

Make an SummarizedExperiment object from an ExpressionSet

Description

makeSummarizedExperimentFromExpressionSet takes a [ExpressionSet](#) object as input and a Range mapping function that maps the features to ranges. It then returns a [SummarizedExperiment](#) object.

Usage

```
makeSummarizedExperimentFromExpressionSet(from,
                                          mapFun=naiveRangeMapper)
```

Arguments

`from` An [ExpressionSet](#) object.

`mapFun` A function which takes an [ExpressionSet](#) object and returns a [GRanges](#), or [GRangesList](#) object which corresponds to the genomic ranges used in the [ExpressionSet](#). The `rownames` of the returned [GRanges](#) are used to match the `featureNames` of the [ExpressionSet](#). [naiveRangeMapper](#).

Value

A [SummarizedExperiment](#) object that corresponds to the input.

See Also

[naiveRangeMapper](#), [probeRangeMapper](#), [geneRangeMapper](#)

Examples

```
library(Biobase)
data(sample.ExpressionSet, package = "Biobase")

# using the naive coercion
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet)

# using probe range mapper
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet, probeRangeMapper)

# using the gene range mapper
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet,
                                          geneRangeMapper("TxDb.Hsapiens.UCSC.hg19.knownGene"))
```

mapCoords-methods

Mapping ranges between sequences

Description

DEPRECATED! Use [mapToTranscripts](#) from the **GenomicFeatures** package or [mapToAlignments](#) from the **GenomicAlignments** package instead.

A method for translating a set of input ranges through a [GRangesList](#) object. Returns a [GenomicRanges](#) object.

The generics for `mapCoords` and `pmapCoords` are defined in the **IRanges** package. A method for translating a set of input ranges through a [GAlignments](#) object is defined and in the **GenomicAlignments** package.

Usage

```
## S4 method for signature 'GenomicRanges,GRangesList'
mapCoords(from, to, ...,
           ignore.strand = TRUE, elt.hits = FALSE)

## S4 method for signature 'GenomicRanges,GRangesList'
pmapCoords(from, to, ...,
            ignore.strand = TRUE, elt.hits = FALSE)
```

Arguments

from	The input ranges to map, usually a GRanges .
to	The alignment between the sequences in from and the sequences in the result, usually a GRangesList .
ignore.strand	logical; When TRUE strand is ignored in overlap operations.
elt.hits	logical; When TRUE, the output includes a metadata column, eltHits, with indices of the inner list elements of to hit by from. Useful for identifying elements of to hit by from. See examples.
...	Arguments passed to other methods. Currently not used.

Details

DEPRECATED! Use [mapToTranscripts](#) from the **GenomicFeatures** package or [mapToAlignments](#) from the **GenomicAlignments** package instead.

Each element in to is taken to represent an alignment of a sequence on a genome. The typical case is a set of transcript models, as might be obtained via `GenomicFeatures::exonsBy`. Each outer list element of the `GRangesList` represents a transcript while each individual element is an exon in the transcript.

`mapCoords` and `pmapCoords` translate the ranges in from relative to the transcript start (i.e., start of all ranges in to). The widths of the individual elements (exons in this example) are concatenated and counting starts at the 5' or 3' end depending on strand. Translated coordinates are only reported for ranges in from that fall completely 'within' ranges in to.

The transcript-centric coordinates are useful, for example, when predicting coding consequences of changes to the genomic sequence.

`mapCoords` maps the i-th element in from to each element in to returning in a many-to-many mapping. In contrast, `pmapCoords` treats the two inputs as parallel vectors and maps the i-th element of from to the i-th element of to returning a maximum of one result per input element.

Value

A `GRanges` object of mapped coordinates with matching data as metadata columns `fromHits` and `toHits`. The ranges in the output `GRanges` are position relative to the outer list element of to; all individual list elements are concatenated and counting starts at the 5' or 3' end depending on strand.

Matching data are the result of calling `findOverlaps` with type 'within' on ranges in from (the query) and the ranges in to (the subject). In the case of `mapCoords` matching can be many-to-one

or one-to-many; one row is reported for each match. For `pmapCoords` matching is one-to-one as the *i*-th element in `from` is only mapped to the *i*-th element in `to`.

When `elt.hits` is `TRUE`, the `eltHits` metadata column includes the index of inner list elements in `to` hit by `from`. In some applications it may be useful to identify the exact list element that was overlapped. These elements can be extracted with the combination of `toHits` (outer list index) and `eltHits` (inner list index).

Author(s)

M. Lawrence and V. Obenchain vobencha@fhcrc.org

See Also

- The generic [mapCoords-methods](#) in the `IRanges` package.
- Additional methods in the `GenomicAlignments` package [mapCoords-methods](#).

Examples

```
## DEPRECATED! See ?mapToTranscripts in the GenomicFeatures package and
## ?mapToAlignments in the GenomicAlignments package.
```

mappingFunctions	<i>Mapping Functions used in <code>makeSummarizedExperimentFromExpressionSet</code></i>
------------------	---

Description

These mapping functions are used in the `mapFun` argument to [makeSummarizedExperimentFromExpressionSet](#).

Usage

```
naiveRangeMapper(from)

probeRangeMapper(from)

geneRangeMapper(txDbPackage, key = "ENTREZID")
```

Arguments

<code>from</code>	An ExpressionSet object.
<code>txDbPackage</code>	A character string with the Transcript Database to use for the mapping.
<code>key</code>	A character string with the Gene key to use for the mapping.

Value

A [GRanges](#) object, with the rownames corresponding to the [featureNames](#) of the [ExpressionSet](#) object.

Examples

```

library(Biobase)
data(sample.ExpressionSet, package = "Biobase")

# using the naive coercion
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet)

# using probe range mapper
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet, probeRangeMapper)

# using the gene range mapper
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet,
                                           geneRangeMapper("TxDb.Hsapiens.UCSC.hg19.knownGene"))

```

nearest-methods

*Finding the nearest genomic range neighbor***Description**

The nearest, precede, follow, distance and distanceToNearest methods for [GenomicRanges](#) objects and subclasses.

Usage

```

## S4 method for signature 'GenomicRanges,GenomicRanges'
precede(x, subject, select=c("arbitrary", "all"), ignore.strand=FALSE)
## S4 method for signature 'GenomicRanges,missing'
precede(x, subject, select=c("arbitrary", "all"), ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges,GenomicRanges'
follow(x, subject, select=c("arbitrary", "all"), ignore.strand=FALSE)
## S4 method for signature 'GenomicRanges,missing'
follow(x, subject, select=c("arbitrary", "all"), ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges,GenomicRanges'
nearest(x, subject, select=c("arbitrary", "all"),
        algorithm=c("nclist", "intervaltree"), ignore.strand=FALSE)
## S4 method for signature 'GenomicRanges,missing'
nearest(x, subject, select=c("arbitrary", "all"),
        algorithm=c("nclist", "intervaltree"), ignore.strand=FALSE)

## S4 method for signature 'GenomicRanges,GenomicRanges'
distanceToNearest(x, subject, algorithm=c("nclist", "intervaltree"),
                 ignore.strand=FALSE, ...)
## S4 method for signature 'GenomicRanges,missing'
distanceToNearest(x, subject, algorithm=c("nclist", "intervaltree"),

```

```
ignore.strand=FALSE, ...)
```

```
## S4 method for signature 'GenomicRanges,GenomicRanges'
distance(x, y, ignore.strand=FALSE, ...)
```

Arguments

x	The query GenomicRanges instance.
subject	The subject GenomicRanges instance within which the nearest neighbors are found. Can be missing, in which case x is also the subject.
y	For the distance method, a GRanges instance. Cannot be missing. If x and y are not the same length, the shortest will be recycled to match the length of the longest.
select	Logic for handling ties. By default, all methods select a single interval (arbitrary for nearest, the first by order in subject for precede, and the last for follow). When select="all" a Hits object is returned with all matches for x. If x does not have a match in subject the x is not included in the Hits object.
algorithm	This argument is passed to findOverlaps , which nearest and distanceToNearest use internally. See ?findOverlaps for more information. Note that it will be removed in BioC 3.3 so please don't use it unless you have a good reason to do so (e.g. troubleshooting).
ignore.strand	A logical indicating if the strand of the input ranges should be ignored. When TRUE, strand is set to '+'.
...	Additional arguments for methods.

Details

- nearest: Performs conventional nearest neighbor finding. Returns an integer vector containing the index of the nearest neighbor range in subject for each range in x. If there is no nearest neighbor NA is returned. For details of the algorithm see the man page in [IRanges](#), [?nearest](#).
- precede: For each range in x, precede returns the index of the range in subject that is directly preceded by the range in x. Overlapping ranges are excluded. NA is returned when there are no qualifying ranges in subject.
- follow: The opposite of precede, follow returns the index of the range in subject that is directly followed by the range in x. Overlapping ranges are excluded. NA is returned when there are no qualifying ranges in subject.
- Orientation and Strand: The relevant orientation for precede and follow is 5' to 3', consistent with the direction of translation. Because positional numbering along a chromosome is from left to right and transcription takes place from 5' to 3', precede and follow can appear to have 'opposite' behavior on the + and - strand. Using positions 5 and 6 as an example, 5 precedes 6 on the + strand but follows 6 on the - strand.

A range with strand * can be compared to ranges on either the + or - strand. Below we outline the priority when ranges on multiple strands are compared. When ignore.strand=TRUE all ranges are treated as if on the + strand.

- x on + strand can match to ranges on both + and * strands. In the case of a tie the first range by order is chosen.

- x on - strand can match to ranges on both - and * strands. In the case of a tie the first range by order is chosen.
- x on * strand can match to ranges on any of +, - or * strands. In the case of a tie the first range by order is chosen.
- distanceToNearest: Returns the distance for each range in x to its nearest neighbor in the subject.
- distance: Returns the distance for each range in x to the range in y. The behavior of distance has changed in Bioconductor 2.12. See the man page ?distance in IRanges for details.

Value

For nearest, precede and follow, an integer vector of indices in subject, or a [Hits](#) if select="all".

For distanceToNearest, a [Hits](#) object with a column for the query index (queryHits), subject index (subjectHits) and the distance between the pair.

For distance, an integer vector of distances between the ranges in x and y.

Author(s)

P. Aboyoun and V. Obenchain <vobencha@fhcrc.org>

See Also

- The [GenomicRanges](#) and [GRanges](#) classes.
- The [Ranges](#) class in the [IRanges](#) package.
- The [Hits](#) class in the [S4Vectors](#) package.
- The [nearest-methods](#) man page in the [IRanges](#) package.
- [findOverlaps-methods](#) for finding just the overlapping ranges.
- The [nearest-methods](#) man page in the [GenomicFeatures](#) package.

Examples

```
## -----
## precede() and follow()
## -----
query <- GRanges("A", IRanges(c(5, 20), width=1), strand="+")
subject <- GRanges("A", IRanges(rep(c(10, 15), 2), width=1),
                        strand=c("+", "+", "-", "-"))
precede(query, subject)
follow(query, subject)

strand(query) <- "-"
precede(query, subject)
follow(query, subject)

## ties choose first in order
query <- GRanges("A", IRanges(10, width=1), c("+", "-", "*"))
subject <- GRanges("A", IRanges(c(5, 5, 5, 15, 15, 15), width=1),
                        rep(c("+", "-", "*"), 2))
```

```

precede(query, subject)
precede(query, rev(subject))

## ignore.strand=TRUE treats all ranges as '+'
precede(query[1], subject[4:6], select="all", ignore.strand=FALSE)
precede(query[1], subject[4:6], select="all", ignore.strand=TRUE)

## -----
## nearest()
## -----
## When multiple ranges overlap an "arbitrary" range is chosen
query <- GRanges("A", IRanges(5, 15))
subject <- GRanges("A", IRanges(c(1, 15), c(5, 19)))
nearest(query, subject)

## select="all" returns all hits
nearest(query, subject, select="all")

## Ranges in 'x' will self-select when 'subject' is present
query <- GRanges("A", IRanges(c(1, 10), width=5))
nearest(query, query)

## Ranges in 'x' will not self-select when 'subject' is missing
nearest(query)

## -----
## distance(), distanceToNearest()
## -----
## Adjacent, overlap, separated by 1
query <- GRanges("A", IRanges(c(1, 2, 10), c(5, 8, 11)))
subject <- GRanges("A", IRanges(c(6, 5, 13), c(10, 10, 15)))
distance(query, subject)

## recycling
distance(query[1], subject)

## zero-width ranges
zw <- GRanges("A", IRanges(4,3))
stopifnot(distance(zw, GRanges("A", IRanges(3,4))) == 0L)
sapply(-3:3, function(i)
  distance(shift(zw, i), GRanges("A", IRanges(4,3))))

query <- GRanges(c("A", "B"), IRanges(c(1, 5), width=1))
distanceToNearest(query, subject)

## distance() with GRanges and TxDb see the
## '?distance,GenomicRanges,TxDb-method' man
## page in the GenomicFeatures package.

```

phicoef

Calculate the "phi coefficient" between two binary variables

Description

The phicoef function calculates the "phi coefficient" between two binary variables.

Usage

```
phicoef(x, y=NULL)
```

Arguments

`x`, `y` Two logical vectors of the same length. If `y` is not supplied, `x` must be a 2x2 integer matrix (or an integer vector of length 4) representing the contingency table of two binary variables.

Value

The "phi coefficient" between the two binary variables. This is a single numeric value ranging from -1 to +1.

Author(s)

H. Pages

References

http://en.wikipedia.org/wiki/Phi_coefficient

Examples

```
set.seed(33)
x <- sample(c(TRUE, FALSE), 100, replace=TRUE)
y <- sample(c(TRUE, FALSE), 100, replace=TRUE)
phicoef(x, y)
phicoef(rep(x, 10), c(rep(x, 9), y))

stopifnot(phicoef(table(x, y)) == phicoef(x, y))
stopifnot(phicoef(y, x) == phicoef(x, y))
stopifnot(phicoef(x, !y) == - phicoef(x, y))
stopifnot(phicoef(x, x) == 1)
```

range-squeezers

*Squeeze the ranges out of a range-based object***Description**

S4 generic functions for squeezing the ranges out of a range-based object.

`granges` returns them as a [GRanges](#) object, `grglist` as a [GRangesList](#) object, and `rglist` as a [RangesList](#) object.

Usage

```
granges(x, use.mcols=FALSE, ...)
grglist(x, use.mcols=FALSE, ...)
rglist(x, use.mcols=FALSE, ...)
```

Arguments

<code>x</code>	A range-based object e.g. a SummarizedExperiment , GAlignments , GAlignmentPairs , or GAlignmentsList object.
<code>use.mcols</code>	TRUE or FALSE (the default). Whether the metadata columns on <code>x</code> (accessible with <code>mcols(x)</code>) should be propagated to the returned object or not.
<code>...</code>	Additional arguments, for use in specific methods.

Details

The [GenomicRanges](#) and [GenomicAlignments](#) packages define and document methods for various types of range-based objects (e.g. for [SummarizedExperiment](#), [GAlignments](#), [GAlignmentPairs](#), and [GAlignmentsList](#) objects). Other Bioconductor packages might as well.

Note that these functions can be seen as a specific kind of *object getters* as well as functions performing coercion. For some objects (e.g. [GAlignments](#)), `as(x, "GRanges")`, `as(x, "GRangesList")`, and `as(x, "RangesList")`, are equivalent to `granges(x, use.mcols=TRUE)`, `grglist(x, use.mcols=TRUE)`, and `rglist(x, use.mcols=TRUE)`, respectively.

Value

A [GRanges](#) object for `granges`.

A [GRangesList](#) object for `grglist`.

A [RangesList](#) object for `rglist`.

If `x` is a vector-like object (e.g. [GAlignments](#)), the returned object is expected to be *parallel* to `x`, that is, the *i*-th element in the output corresponds to the *i*-th element in the input. If `x` has names on it, they're propagated to the returned object. If `use.mcols` is TRUE and `x` has metadata columns on it (accessible with `mcols(x)`), they're propagated to the returned object.

Author(s)

H. Pages

See Also

- [GRanges](#) and [GRangesList](#) objects.
- [RangesList](#) objects in the **IRanges** package.
- [SummarizedExperiment](#) objects.
- [GAlignments](#), [GAlignmentPairs](#), and [GAlignmentsList](#) objects in the **GenomicAlignments** package.

Examples

```
## See ?GAlignments in the GenomicAlignments package for some
## examples.
```

 setops-methods

Set operations on GRanges and GRangesList objects

Description

Performs set operations on [GRanges](#) and [GRangesList](#) objects.

Usage

```
## Set operations
## S4 method for signature 'GRanges,GRanges'
union(x, y, ignore.strand=FALSE, ...)
## S4 method for signature 'GRanges,GRanges'
intersect(x, y, ignore.strand=FALSE, ...)
## S4 method for signature 'GRanges,GRanges'
setdiff(x, y, ignore.strand=FALSE, ...)

## Parallel set operations
## S4 method for signature 'GRanges,GRanges'
punion(x, y, fill.gap=FALSE, ignore.strand=FALSE, ...)
## S4 method for signature 'GRanges,GRanges'
pintersect(x, y, resolve.empty=c("none", "max.start", "start.x"), ignore.strand=FALSE, ...)
## S4 method for signature 'GRanges,GRanges'
psetdiff(x, y, ignore.strand=FALSE, ...)
```

Arguments

`x, y` For `union`, `intersect`, `setdiff`, `pgap`: `x` and `y` must both be [GRanges](#) objects. For `punion`: one of `x` or `y` must be a [GRanges](#) object, the other one can be a [GRanges](#) or [GRangesList](#) object. For `pintersect`: one of `x` or `y` must be a [GRanges](#) object, the other one can be a [GRanges](#) or [GRangesList](#) object.

For `psetdiff`: `x` and `y` can be any combination of [GRanges](#) and/or [GRangesList](#) objects, with the exception that if `x` is a [GRangesList](#) object then `y` must be a [GRangesList](#) too.

In addition, for the "parallel" operations, `x` and `y` must be of equal length (i.e. `length(x) == length(y)`).

<code>fill.gap</code>	Logical indicating whether or not to force a union by using the rule <code>start = min(start(x), start(y))</code> .
<code>resolve.empty</code>	One of "none", "max.start", or "start.x" denoting how to handle ambiguous empty ranges formed by intersections. "none" - throw an error if an ambiguous empty range is formed, "max.start" - associate the maximum start value with any ambiguous empty range, and "start.x" - associate the start value of <code>x</code> with any ambiguous empty range. (See pintersect for the definition of an ambiguous range.)
<code>ignore.strand</code>	For set operations: If set to TRUE, then the strand of <code>x</code> and <code>y</code> is set to "*" prior to any computation. For parallel set operations: If set to TRUE, the strand information is ignored in the computation and the result has the strand information of <code>x</code> .
...	Further arguments to be passed to or from other methods.

Details

The `pintersect` methods involving [GRanges](#) and/or [GRangesList](#) objects use the triplet (sequence name, range, strand) to determine the element by element intersection of features, where a strand value of "*" is treated as occurring on both the "+" and "-" strand.

The `psetdiff` methods involving [GRanges](#) and/or [GRangesList](#) objects use the triplet (sequence name, range, strand) to determine the element by element set difference of features, where a strand value of "*" is treated as occurring on both the "+" and "-" strand.

Value

For `union`, `intersect`, `setdiff`, and `pgap`: a [GRanges](#).

For `punion` and `pintersect`: when `x` or `y` is not a [GRanges](#) object, an object of the same class as this non-[GRanges](#) object. Otherwise, a [GRanges](#) object.

For `psetdiff`: either a [GRanges](#) object when both `x` and `y` are [GRanges](#) objects, or a [GRangesList](#) object when `y` is a [GRangesList](#) object.

Author(s)

P. Aboyoun and H. Pages

See Also

[setops-methods](#), [GRanges-class](#), [GRangesList-class](#), [findOverlaps-methods](#)

Examples

```

## -----
## A. SET OPERATIONS
## -----

x <- GRanges("chr1", IRanges(c(2, 9) , c(7, 19)), strand=c("+", "-"))
y <- GRanges("chr1", IRanges(5, 10), strand="-")

union(x, y)
union(x, y, ignore.strand=TRUE)

intersect(x, y)
intersect(x, y, ignore.strand=TRUE)

setdiff(x, y)
setdiff(x, y, ignore.strand=TRUE)

## -----
## B. PARALLEL SET OPERATIONS
## -----

## Not run:
punion(x, shift(x, 7)) # will fail

## End(Not run)
punion(x, shift(x, 7), fill.gap=TRUE)

pintersect(x, shift(x, 6))
## Not run:
pintersect(x, shift(x, 7)) # will fail

## End(Not run)
pintersect(x, shift(x, 7), resolve.empty="max.start")

psetdiff(x, shift(x, 7))

## -----
## C. MORE EXAMPLES
## -----

## GRanges object:
gr <- GRanges(seqnames=c("chr2", "chr1", "chr1"),
              ranges=IRanges(1:3, width = 12),
              strand=Rle(strand(c("-", "x", "-"))))

## GRangesList object
gr1 <- GRanges(seqnames="chr2",
              ranges=IRanges(3, 6))
gr2 <- GRanges(seqnames=c("chr1", "chr1"),
              ranges=IRanges(c(7,13), width = 3),
              strand=c("+", "-"))
gr3 <- GRanges(seqnames=c("chr1", "chr2"),

```

```

        ranges=IRanges(c(1, 4), c(3, 9)),
        strand=c("-", "-"))
grlist <- GRangesList(gr1=gr1, gr2=gr2, gr3=gr3)

## Parallel intersection of a GRanges and a GRangesList object
pintersect(gr, grlist)
pintersect(grlist, gr)

## Parallel set difference of a GRanges and a GRangesList object
psetdiff(gr, grlist)

## Parallel set difference of two GRangesList objects
psetdiff(grlist, shift(grlist, 3))

```

strand-utils

Strand utilities

Description

Some useful strand methods.

Usage

```

## S4 method for signature 'missing'
strand(x)
## S4 method for signature 'character'
strand(x)
## S4 method for signature 'factor'
strand(x)
## S4 method for signature 'integer'
strand(x)
## S4 method for signature 'logical'
strand(x)
## S4 method for signature 'Rle'
strand(x)
## S4 method for signature 'DataTable'
strand(x)
## S4 replacement method for signature 'DataTable,ANY'
strand(x) <- value

```

Arguments

x	The object from which to obtain a strand factor, can be missing.
value	Replacement value for the strand.

Details

If `x` is missing, returns an empty factor with the "standard strand levels" i.e. `+`, `-`, and `*`.

If `x` is a character vector or factor, it is coerced to a factor with the levels listed above. NA values in `x` are not accepted.

If `x` is an integer vector, it is coerced to a factor with the levels listed above. `1`, `-1`, and NA values in `x` are mapped to the `+`, `-`, and `*` levels respectively.

If `x` is a logical vector, it is coerced to a factor with the levels listed above. `FALSE`, `TRUE`, and NA values in `x` are mapped to the `+`, `-`, and `*` levels respectively.

If `x` is a character-, factor-, integer-, or logical-[Rle](#), it is transformed with `runValue(x) <- strand(runValue(x))` and returned.

If `x` inherits from `DataTable`, the "strand" column is passed thru `strand()` and returned. If `x` has no "strand" column, this return value is populated with `*`s.

Value

A factor (or factor-[Rle](#)) with the "standard strand levels" (i.e. `+`, `-`, and `*`) and no NAs.

Author(s)

Michael Lawrence and H. Pages

See Also

[strand](#)

Examples

```
strand()

x1 <- c("-", "*", "*", "+", "-", "*")
x2 <- factor(c("-", "-", "+", "-"))
x3 <- c(-1L, NA, NA, 1L, -1L, NA)
x4 <- c(TRUE, NA, NA, FALSE, TRUE, NA)

strand(x1)
strand(x2)
strand(x3)
strand(x4)

strand(Rle(x1))
strand(Rle(x2))
strand(Rle(x3))
strand(Rle(x4))

strand(DataFrame(score=2:-3))
strand(DataFrame(score=2:-3, strand=x3))
strand(DataFrame(score=2:-3, strand=Rle(x3)))

## Sanity checks:
```

```

target <- strand(x1)
stopifnot(identical(target, strand(x3)))
stopifnot(identical(target, strand(x4)))

stopifnot(identical(Rle(strand(x1)), strand(Rle(x1))))
stopifnot(identical(Rle(strand(x2)), strand(Rle(x2))))
stopifnot(identical(Rle(strand(x3)), strand(Rle(x3))))
stopifnot(identical(Rle(strand(x4)), strand(Rle(x4))))

```

SummarizedExperiment-class

SummarizedExperiment instances

Description

The SummarizedExperiment class is a matrix-like container where rows represent ranges of interest (as a [GRanges](#) or [GRangesList](#)-class) and columns represent samples (with sample data summarized as a [DataFrame](#)-class). A SummarizedExperiment contains one or more assays, each represented by a matrix-like object of numeric or other mode.

Usage

```

## Constructors

SummarizedExperiment(assays, ...)
## S4 method for signature 'SimpleList'
SummarizedExperiment(assays, rowRanges = GRangesList(),
  colData = DataFrame(), exptData = SimpleList(), ...,
  verbose = FALSE)
## S4 method for signature 'missing'
SummarizedExperiment(assays, ...)
## S4 method for signature 'list'
SummarizedExperiment(assays, ...)
## S4 method for signature 'matrix'
SummarizedExperiment(assays, ...)

## Accessors

assayNames(x, ...)
assayNames(x, ...) <- value
assays(x, ..., withDimnames=TRUE)
assays(x, ..., withDimnames=TRUE) <- value
assay(x, i, ...)
assay(x, i, ...) <- value
rowRanges(x, ...)
rowRanges(x, ...) <- value

```



```
colData(x, ...)
colData(x, ...) <- value
exptData(x, ...)
exptData(x, ...) <- value
## S4 method for signature 'SummarizedExperiment'
dim(x)
## S4 method for signature 'SummarizedExperiment'
dimnames(x)
## S4 replacement method for signature 'SummarizedExperiment,NULL'
dimnames(x) <- value
## S4 replacement method for signature 'SummarizedExperiment,list'
dimnames(x) <- value

## colData access

## S4 method for signature 'SummarizedExperiment'
x$name
## S4 replacement method for signature 'SummarizedExperiment,ANY'
x$name <- value
## S4 method for signature 'SummarizedExperiment,ANY,missing'
x[[i, j, ...]]
## S4 replacement method for signature 'SummarizedExperiment,ANY,missing,ANY'
x[[i, j, ...]] <- value

## rowRanges access
## see 'GRanges compatibility', below

## Subsetting

## S4 method for signature 'SummarizedExperiment'
x[i, j, ..., drop=TRUE]
## S4 replacement method for signature 'SummarizedExperiment,ANY,ANY,SummarizedExperiment'
x[i, j] <- value
## S4 method for signature 'SummarizedExperiment'
subset(x, subset, select, ...)

## Combining

## S4 method for signature 'SummarizedExperiment'
cbind(..., deparse.level=1)
## S4 method for signature 'SummarizedExperiment'
rbind(..., deparse.level=1)

## Coercion

## S4 method for signature 'SummarizedExperiment'
updateObject(object, ..., verbose=FALSE)
## S4 method for signature 'ExpressionSet,SummarizedExperiment'
```

```

coerce(from, to = "SummarizedExperiment", strict = TRUE)
## S4 method for signature 'SummarizedExperiment,ExpressionSet'
coerce(from, to = "ExpressionSet", strict = TRUE)

```

Arguments

assays	A list or SimpleList of matrix elements, or a matrix. All elements of the list must have the same dimensions, and dimension names (if present) must be consistent across elements and with the row names of rowRanges and colData.
rowRanges	A GRanges or GRangesList instance describing the ranges of interest. Row names, if present, become the row names of the SummarizedExperiment. The length of the GRanges or the GRangesList must equal the number of rows of the matrices in assays.
colData	An optional DataFrame describing the samples. Row names, if present, become the column names of the SummarizedExperiment.
exptData	An optional SimpleList of arbitrary content describing the overall experiment.
...	For SummarizedExperiment, S4 methods list and matrix, arguments identical to those of the SimpleList method. For assay, ... may contain withDimnames, which is forwarded to assays. For cbind, rbind, ... contains SummarizedExperiment objects to be combined. For other accessors, ignored.
verbose	A logical(1) indicating whether messages about data coercion during construction should be printed.
x, object	An instance of SummarizedExperiment-class.
i, j	For assay, assay<-, i is a integer or numeric scalar; see 'Details' for additional constraints. For [, SummarizedExperiment, [, SummarizedExperiment<-, i, j are instances that can act to subset the underlying rowRanges, colData, and matrix elements of assays. For [[, SummarizedExperiment, [[<-, SummarizedExperiment, i is a scalar index (e.g., character(1) or integer(1)) into a column of colData.
subset	An expression which, when evaluated in the context of rowRanges(x), is a logical vector indicating elements or rows to keep: missing values are taken as false.
select	An expression which, when evaluated in the context of colData(x), is a logical vector indicating elements or rows to keep: missing values are taken as false.
name	A symbol representing the name of a column of colData.
withDimnames	A logical(1), indicating whether dimnames should be applied to extracted assay elements. Setting withDimnames=FALSE increases the speed and memory efficiency with which assays are extracted. withDimnames=TRUE in the getter assays<- allows efficient complex assignments (e.g., updating names of assays, names(assays(x, withDimnames=FALSE)) = ... is more efficient than names(assays(x)) = ...); it does not influence actual assignment of dimnames to assays.

drop	A logical(1), ignored by these methods.
value	An instance of a class specified in the S4 method signature or as outlined in ‘Details’.
deparse.level	See <code>?base::cbind</code> for a description of this argument.
from	the object to be coerced
to	the class to coerce to
strict	logical flag. If ‘TRUE’, the returned object must be strictly from the target class.

Details

The `SummarizedExperiment` class is meant for numeric and other data types derived from a sequencing experiment. The structure is rectangular like a `matrix`, but with additional annotations on the rows and columns, and with the possibility to manage several assays simultaneously.

The rows of a `SummarizedExperiment` instance represent ranges (in genomic coordinates) of interest. The ranges of interest are described by a `GRanges`-class or a `GRangesList`-class instance, accessible using the `rowRanges` function, described below. The `GRanges` and `GRangesList` classes contains sequence (e.g., chromosome) name, genomic coordinates, and strand information. Each range can be annotated with additional data; this data might be used to describe the range or to summarize results (e.g., statistics of differential abundance) relevant to the range. Rows may or may not have row names; they often will not.

Each column of a `SummarizedExperiment` instance represents a sample. Information about the samples are stored in a `DataFrame`-class, accessible using the function `colData`, described below. The `DataFrame` must have as many rows as there are columns in the `SummarizedExperiment`, with each row of the `DataFrame` providing information on the sample in the corresponding column of the `SummarizedExperiment`. Columns of the `DataFrame` represent different sample attributes, e.g., tissue of origin, etc. Columns of the `DataFrame` can themselves be annotated (via the `mcols` function). Column names typically provide a short identifier unique to each sample.

A `SummarizedExperiment` can also contain information about the overall experiment, for instance the lab in which it was conducted, the publications with which it is associated, etc. This information is stored as a `SimpleList`-class, accessible using the `exptData` function. The form of the data associated with the experiment is left to the discretion of the user.

The `SummarizedExperiment` is appropriate for matrix-like data. The data are accessed using the `assays` function, described below. This returns a `SimpleList`-class instance. Each element of the list must itself be a matrix (of any mode) and must have dimensions that are the same as the dimensions of the `SummarizedExperiment` in which they are stored. Row and column names of each matrix must either be `NULL` or match those of the `SummarizedExperiment` during construction. It is convenient for the elements of `SimpleList` of assays to be named.

The `SummarizedExperiment` class has the following slots; this detail of class structure is not relevant to the user.

- `exptData` A `SimpleList`-class instance containing information about the overall experiment.
- `rowData` A `GRanges`-class instance defining the ranges of interest and associated metadata. **WARNING:** The accessor for this slot is `rowRanges`, not `rowData`!
- `colData` A `DataFrame`-class instance describing the samples and associated metadata.
- `assays` A `SimpleList`-class instance, each element of which is a matrix summarizing data associated with the corresponding range and sample.

Constructor

Instances are constructed using the `SummarizedExperiment` function with arguments outlined above.

Coercion

Package version 1.9.59 introduced a new way of representing ‘assays’. If you have a serialized instance `x` of a `SummarizedExperiment` (e.g., from using the `save` function with a version of `GenomicRanges` prior to 1.9.59), it should be updated by invoking `x <- updateObject(x)`.

`as(from, "SummarizedExperiment")`: Creates a `SummarizedExperiment` object from a `ExpressionSet` object.

`as(from, "ExpressionSet")`: Creates a `ExpressionSet` object from a `SummarizedExperiment` object.

The following data mappings are used for coercion between `ExpressionSet` and `SummarizedExperiment`.

```
assayData assays
featureData rowData
phenoData colData
experimentData, annotation, protocolData colData
```

If the `SummarizedExperiment` being coerced uses `GRanges` to store its range data that data will be included in the `featureData` of the `ExpressionSet`.

Because `ExpressionSet` objects require an assay named ‘`exprs`’ if the `SummarizedExperiment` object being coerced does not have an assay named ‘`exprs`’ the first assay will be renamed and a warning will be issued.

Accessors

In the following code snippets, `x` is a `SummarizedExperiment` instance.

`assays(x)`, `assays(x) <- value`: Get or set the assays. `value` is a list or `SimpleList`, each element of which is a matrix with the same dimensions as `x`.

`assay(x, i)`, `assay(x, i) <- value`: A convenient alternative (to `assays(x)[[i]]`, `assays(x)[[i]] <- value`) to get or set the `i`th (default first) assay element. `value` must be a matrix of the same dimension as `x`, and with dimension names `NULL` or consistent with those of `x`.

`assayNames(x)`, `assayNames(x) <- value`: Get or set the names of `assay()` elements.

`rowRanges(x)`, `rowRanges(x) <- value`: Get or set the row data. `value` is a `GenomicRanges` instance. Row names of `value` must be `NULL` or consistent with the existing row names of `x`.

`colData(x)`, `colData(x) <- value`: Get or set the column data. `value` is a `DataFrame` instance. Row names of `value` must be `NULL` or consistent with the existing column names of `x`.

`exptData(x)`, `exptData(x) <- value`: Get or set the experiment data. `value` is a list or `SimpleList` instance, with arbitrary content.

`dim(x)`: Get the dimensions (ranges x samples) of the `SummarizedExperiment`.

`dimnames(x)`, `dimnames(x) <- value`: Get or set the dimension names. `value` is usually a list of length 2, containing elements that are either `NULL` or vectors of appropriate length for the corresponding dimension. `value` can be `NULL`, which removes dimension names. This method implies that `rownames`, `rownames<-`, `colnames`, and `colnames<-` are all available.

GRanges compatibility (rowRanges access)

Many `GRanges`-class and `GRangesList`-class operations are supported on ‘SummarizedExperiment’ and derived instances, using `rowRanges`.

Supported operations include: `compare`, `countOverlaps`, `coverage`, `disjointBins`, `distance`, `distanceToNearest`, `duplicated`, `end`, `end<-`, `findOverlaps`, `flank`, `follow`, `granges`, `isDisjoint`, `match`, `mcols`, `mcols<-`, `narrow`, `nearest`, `order`, `overlapsAny`, `precede`, `ranges`, `ranges<-`, `rank`, `resize`, `restrict`, `seqinfo`, `seqinfo<-`, `seqnames`, `shift`, `sort`, `split`, `relistToClass`, `start`, `start<-`, `strand`, `strand<-`, `subsetByOverlaps`, `width`, `width<-`.

Not all `GRanges`-class operations are supported, because they do not make sense for ‘SummarizedExperiment’ objects (e.g., `length`, `name`, `as.data.frame`, `c`, `splitAsList`), involve non-trivial combination or splitting of rows (e.g., `disjoin`, `gaps`, `reduce`, `unique`), or have not yet been implemented (`Ops`, `map`, `window`, `window<-`).

Subsetting

In the code snippets below, `x` is a `SummarizedExperiment` instance.

`x[i,j], x[[i,j]] <- value`: Create or replace a subset of `x`. `i`, `j` can be numeric, logical, character, or missing. `value` must be a `SummarizedExperiment` instance with dimensions, dimension names, and assay elements consistent with the subset `x[i,j]` being replaced.

`subset(x, subset, select)`: Create a subset of `x` using an expression `subset` referring to columns of `rowRanges(x)` (including ‘`seqnames`’, ‘`start`’, ‘`end`’, ‘`width`’, ‘`strand`’, and `names(mcols(x))`) and / or `select` referring to column names of `colData(x)`.

Additional subsetting accessors provide convenient access to `colData` columns

`x$name, x[[name]] <- value` Access or replace column name in `x`.

`x[[i, ...]], x[[[i, ...]]] <- value` Access or replace column `i` in `x`.

Combining

In the code snippets below, `...` are `SummarizedExperiment` instances to be combined.

`cbind(...)`, `rbind(...)`: `cbind` combines objects with identical ranges (`rowRanges`) but different samples (columns in assays). The `colnames` in `colData` must match or an error is thrown. Duplicate columns of `mcols(rowRanges(SummarizedExperiment))` must contain the same data. Data in assays are combined by name matching; if all names are `NULL` matching is by position. A mixture of names and `NULL` throws an error.

`rbind` combines objects with different ranges (`rowRanges`) and the same subjects (columns in assays). Duplicate columns of `colData` must contain the same data.

`exptData` from all objects are combined into a `SimpleList` with no name checking.

Implementation and Extension

This section contains advanced material meant for package developers.

`SummarizedExperiment` is implemented as an S4 class, and can be extended in the usual way, using `contains="SummarizedExperiment"` in the new class definition.

In addition, the representation of the assays slot of `SummarizedExperiment` is as a virtual class `Assays`. This allows derived classes (`contains="Assays"`) to easily implement alternative requirements for the assays, e.g., backed by file-based storage like `NetCDF` or the `ff` package, while re-using the existing `SummarizedExperiment` class without modification. The requirements on `Assays` are list-like semantics (e.g., `sapply`, `[]` subsetting, `names`) with elements having matrix- or array-like semantics (e.g., `dim`, `dimnames`). These requirements can be made more precise if developers express interest.

The current assays slot is implemented as a reference class that has copy-on-change semantics. This means that modifying non-assay slots does not copy the (large) assay data, and at the same time the user is not surprised by reference-based semantics. Updates to non-assay slots are very fast; updating the assays slot itself can be 5x or more faster than with an S4 instance in the slot. One useful technique when working with assay or assays function is use of the `withDimnames=FALSE` argument, which benefits speed and memory use by not copying `dimnames` from the row- and `colData` elements to each assay.

In a little more detail, a small reference class hierarchy (not exported from the `GenomicRanges` name space) defines a reference class `ShallowData` with a single field `data` of type `ANY`, and a derived class `ShallowSimpleListAssays` that specializes the type of data as `SimpleList`, and `contains=c("ShallowData", "Assays")`. The assays slot contains an instance of `ShallowSimpleListAssays`. Invoking `assays()` on a `SummarizedExperiment` re-dispatches from the assays slot to retrieve the `SimpleList` from the field of the reference class. This was achieved by implementing a generic (not exported) `value(x, name, ...)`, with a method implemented on `SummarizedExperiment` that retrieves a slot when `name` is a slot containing an S4 object in `x`, and a field when `name` is a slot containing a `ShallowData` instance in `x`. Copy-on-change semantics is maintained by implementing the `clone` method (`clone` methods are supposed to do a deep copy, update methods a shallow copy; the `clone` generic is introduced, and not exported, in the `GenomicRanges` package). The ‘getter’ and ‘setter’ code for methods implemented on `SummarizedExperiment` use `value` for slot access, and `clone` for replacement. This makes it easy to implement `ShallowData` instances for other slots if the need arises.

Author(s)

Martin Morgan, mtmorgan@fhcrc.org

See Also

[GRanges](#), [DataFrame](#), [SimpleList](#),

Examples

```
nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(50, 150)),
                     IRanges(floor(runif(200, 1e5, 1e6)), width=100),
                     strand=sample(c("+", "-"), 200, TRUE))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
sset <- SummarizedExperiment(assays=SimpleList(counts=counts),
                             rowRanges=rowRanges, colData=colData)
sset
```

```

assayNames(sset)
assays(sset) <- endoapply(assays(sset), asinh)
head(assay(sset))

sset[, sset$Treatment == "ChIP"]

## cbind combines objects with the same ranges and different samples.
se1 <- sset
se2 <- se1[,1:3]
colnames(se2) <- letters[1:ncol(se2)]
cmb1 <- cbind(se1, se2)

## rbind combines objects with the same samples and different ranges.
se1 <- sset
se2 <- se1[1:50,]
rownames(se2) <- letters[1:nrow(se2)]
cmb2 <- rbind(se1, se2)

```

tile-methods

Tile a GenomicRanges object

Description

tile method for **GenomicRanges**. Partitions each range into a set of tiles. Tiles are defined in terms of their number or width.

Usage

```
## S4 method for signature 'GenomicRanges'
tile(x, n, width)
```

Arguments

x	A GenomicRanges object, like a GRanges .
n	The number of tiles to generate. See ?tile in the IRanges package for more information about this argument.
width	The (maximum) width of each tile. See ?tile in the IRanges package for more information about this argument.

Details

Splits x into a **GRangesList**, each element of which corresponds to a tile, or partition, of x. Specify the tile geometry with either n or width (not both). Passing n creates n tiles of approximately equal width, truncated by sequence end, while passing width tiles the region with ranges of the given width, again truncated by sequence end.

Value

A GRangesList object, each element of which corresponds to a tile.

Author(s)

M. Lawrence

See Also

[tile](#) in the **IRanges** package.

Examples

```
gr <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=11),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))

# split every range in half
tiles <- tile(gr, n = 2L)
stopifnot(all(elementLengths(tiles) == 2L))

# split ranges into subranges of width 2
# odd width ranges must contain one subrange of width 1
tiles <- tile(gr, width = 2L)
stopifnot(all(all(width(tiles) %in% c(1L, 2L))))
```

tileGenome

Put (virtual) tiles on a given genome

Description

tileGenome returns a set of genomic regions that form a partitioning of the specified genome. Each region is called a "tile".

Usage

```
tileGenome(seqlengths, ntile, tilewidth, cut.last.tile.in.chrom=FALSE)
```

Arguments

seqlengths Either a named numeric vector of chromosome lengths or a [Seqinfo](#) object. More precisely, if a named numeric vector, it must have a length ≥ 1 , cannot contain NAs or negative values, and cannot have duplicated names. If a [Seqinfo](#) object, then it's first replaced with the vector of sequence lengths stored in the object (extracted from the object with the [seqlengths](#) getter), then the restrictions described previously apply to this vector.

<code>ntile</code>	The number of tiles to generate.
<code>tilewidth</code>	The desired tile width. The effective tile width might be slightly different but is guaranteed to never be more than the desired width.
<code>cut.last.tile.in.chrom</code>	Whether or not to cut the last tile in each chromosome. This is set to FALSE by default. Can be set to TRUE only when <code>tilewidth</code> is specified. In that case, a tile will never overlap with more than 1 chromosome and a GRanges object is returned with one element (i.e. one genomic range) per tile.

Value

If `cut.last.tile.in.chrom` is FALSE (the default), a [GRangesList](#) object with one list element per tile, each of them containing a number of genomic ranges equal to the number of chromosomes it overlaps with. Note that when the tiles are small (i.e. much smaller than the chromosomes), most of them only overlap with a single chromosome.

If `cut.last.tile.in.chrom` is TRUE, a [GRanges](#) object with one element (i.e. one genomic range) per tile.

Author(s)

H. Pages, based on a proposal by Martin Morgan

See Also

- [GRangesList](#) and [GRanges](#) objects.
- [Seqinfo](#) objects and the `seqlengths` getter.
- [IntegerList](#) objects.
- [Views](#) objects.
- [coverage,GenomicRanges-method](#) for computing the coverage of a [GRanges](#) object.

Examples

```
## -----
## A. WITH A TOY GENOME
## -----

seqlengths <- c(chr1=60, chr2=20, chr3=25)

## Create 5 tiles:
tiles <- tileGenome(seqlengths, ntile=5)
tiles
elementLengths(tiles) # tiles 3 and 4 contain 2 ranges

width(tiles)
## Use sum() on this IntegerList object to get the effective tile
## widths:
sum(width(tiles)) # each tile covers exactly 21 genomic positions

## Create 9 tiles:
```

```

tiles <- tileGenome(seqlengths, ntile=9)
elementLengths(tiles) # tiles 6 and 7 contain 2 ranges

table(sum(width(tiles))) # some tiles cover 12 genomic positions,
                        # others 11

## Specify the tile width:
tiles <- tileGenome(seqlengths, tilewidth=20)
length(tiles) # 6 tiles
table(sum(width(tiles))) # effective tile width is <= specified

## Specify the tile width and cut the last tile in each chromosome:
tiles <- tileGenome(seqlengths, tilewidth=24,
                  cut.last.tile.in.chrom=TRUE)
tiles
width(tiles) # each tile covers exactly 24 genomic positions, except
             # the last tile in each chromosome

## Partition a genome by chromosome ("natural partitioning"):
tiles <- tileGenome(seqlengths, tilewidth=max(seqlengths),
                  cut.last.tile.in.chrom=TRUE)
tiles # one tile per chromosome

## sanity check
stopifnot(all.equal(setNames(end(tiles), seqnames(tiles)), seqlengths))

## -----
## B. WITH A REAL GENOME
## -----

library(BSgenome.Scerevisiae.UCSC.sacCer2)
tiles <- tileGenome(seqinfo(Scerevisiae), ntile=20)
tiles

tiles <- tileGenome(seqinfo(Scerevisiae), tilewidth=50000,
                  cut.last.tile.in.chrom=TRUE)
tiles

## -----
## C. AN APPLICATION: COMPUTE THE BINNED AVERAGE OF A NUMERICAL VARIABLE
##   DEFINED ALONG A GENOME
## -----

## 1. When the variable is stored in a named RleList object
## -----

## In Bioconductor, a variable defined along a genome is typically
## represented as a named RleList object with one list element per
## chromosome. Let's create such a variable:

library(BSgenome.Scerevisiae.UCSC.sacCer2)
set.seed(22)
my_var1 <- RleList(

```

```

    lapply(seqlengths(Scerevisiae),
           function(len) Rle(sample(-10:10, len, replace=TRUE))),
    compress=FALSE)
my_var1

## In some applications, there is sometimes the need to compute the
## average of 'my_var1' for each genomic region in a set of predefined
## fixed-width regions (sometimes called "bins"). Let's use
## tileGenome() to create such a set of bins:

bins1 <- tileGenome(seqinfo(Scerevisiae), tilewidth=100,
                    cut.last.tile.in.chrom=TRUE)

## We define the following function to compute the binned average of a
## numerical variable defined along a genome.
## Arguments:
## 'bins': a GRanges object representing the genomic bins.
##         Typically obtained by calling tileGenome() with
##         'cut.last.tile.in.chrom=TRUE'.
## 'numvar': a named RleList object representing a numerical
##            variable defined along the genome covered by 'bins', which
##            is the genome described by 'seqinfo(bins)'.
## 'mcolname': the name to give to the metadata column that will
##             contain the binned average in the returned object.
## Returns 'bins' with an additional metadata column named 'mcolname'
## containing the binned average.

binnedAverage <- function(bins, numvar, mcolname)
{
  stopifnot(is(bins, "GRanges"))
  stopifnot(is(numvar, "RleList"))
  stopifnot(identical(seqlevels(bins), names(numvar)))

  ## A version of viewMeans() that pads "out of limits" views with zeros.
  viewMeans2 <- function(v) {
    means <- viewMeans(v)
    w0 <- width(v)
    w1 <- width(trim(v))
    means <- means * w1 / w0
    means[w0 != 0L & w1 == 0L] <- 0
    means
  }

  bins_per_chrom <- split(ranges(bins), seqnames(bins))
  means_list <- lapply(names(numvar),
                      function(seqname) {
                        v <- Views(numvar[[seqname]], bins_per_chrom[[seqname]])
                        viewMeans2(v)
                      })
  new_mcol <- unsplit(means_list, as.factor(seqnames(bins)))
  mcols(bins)[[mcolname]] <- new_mcol
  bins
}

```

```

## Compute the binned average for 'my_var1':

bins1 <- binnedAverage(bins1, my_var1, "binned_var1")
bins1

## 2. When the variable is stored in a metadata column of a disjoint
## GRanges object
## -----

## A GRanges object is said to be disjoint if it contains ranges
## that do not overlap with each other. This can be tested with the
## isDisjoint() function. For example, the GRanges object returned
## by tileGenome() is always guaranteed to be disjoint:

stopifnot(isDisjoint(bins1))

## In addition to named RleList objects, the metadata columns of a
## disjoint GRanges object can also be seen as variables defined
## along a genome. An obvious example is the "binned_var1" metadata
## column in 'bins1'. Another example is the "score" metadata column
## in the following GRanges object:

x2 <- GRanges("chrI",
              IRanges(c(1, 211, 291), c(150, 285, 377)),
              score=c(0.4, 8, -10),
              seqinfo=seqinfo(Scerevisiae))
x2

## If we consider the score to be zero in the genomic regions not
## covered by 'x2', then the "score" metadata column represents a
## variable defined along the genome.

## Turning the score variable into a named RleList representation
## can be done by computing the weighted coverage of 'x2':

score <- coverage(x2, weight="score")
score

## Now we can pass 'score' to binnedAverage() to compute the average
## score per bin:

bins1 <- binnedAverage(bins1, score, "binned_score")
bins1

## With bigger bins:

bins2 <- tileGenome(seqinfo(x2), tilewidth=50000,
                  cut.last.tile.in.chrom=TRUE)
bins2 <- binnedAverage(bins2, score, "binned_score")
bins2

## Note that the binned variables in 'bins1' and 'bins2' can be

```

```
## turned back into named RleList objects:

binned_var1 <- coverage(bins1, weight="binned_var1")
stopifnot(all.equal(mean(binned_var1), mean(my_var1)))

binned_score <- coverage(bins2, weight="binned_score")
stopifnot(all.equal(mean(binned_score), mean(score)))

## Not surprisingly, the "binned" variables are much more compact in
## memory than the original variables (they contain much less runs):

object.size(binned_var1)
object.size(my_var1)
```

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